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TRANSMITTAL LETTER TO THE UNITED STATES
DESIGNATED/ELECTED OFFICE (DO/EO/US)
CONCERNING A FILING UNDER 35 U.S.C. 371

016786/0215

U.S. APPLICATION NO. (If known, see 37 C.F.R. 1.5)

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INTERNATIONAL APPLICATION NO.

PCT/AU97/00436

INTERNATIONAL FILING DATE

July 9, 1997

PRIORITY DATE CLAIMED

July 6, 1996

TITLE OF INVENTION

NEUROACTIVE PEPTIDE

APPLICANT(S) FOR DO/EO/US

Frederick A. O. MENDELSON, Siew Yeen CHAI, Ingrid MOELLER, Peter G. ALFRED, Ian A. SMITH, and Rebecca A. LEW

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. ☒ This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371.
 2. ☐ This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. 371.
 3. ☐ This express request to begin national examination procedures (35 U.S.C. 371(f)) at any time rather than delay examination until the expiration of the applicable time limit set in 35 U.S.C. 371(b) and PCT Articles 22 and 39(1).
 4. ☒ A proper Demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date.
 5. ☒ A copy of the International Application as filed (35 U.S.C. 371(c)(2))
 - a. ☒ is transmitted herewith (required only if not transmitted by the International Bureau).
 - b. ☐ has been transmitted by the International Bureau.
 - c. ☐ is not required, as the application was filed in the United States Receiving Office (RO/US).
 6. ☐ A translation of the International Application into English (35 U.S.C. 371 (c)(2)).
 7. ☒ Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(3))
 - a. ☐ are transmitted herewith (required only if not transmitted by the International Bureau).
 - b. ☐ have been transmitted by the International Bureau.
 - c. ☐ have not been made; however, the time limit for making such amendments has NOT expired.
 - d. ☒ have not been made and will not be made.
 8. ☐ A translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).
 9. ☐ An oath or declaration of the inventor(s) (35 U.S.C. 371(c)(4)).
 10. ☐ A translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)).
- Items 11. to 16. below concern other document(s) or information included:**
11. ☒ An Information Disclosure Statement under 37 CFR 1.97 and 1.98.
 12. ☐ An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
 13. ☒ A **FIRST** preliminary amendment.
☐ A **SECOND** or **SUBSEQUENT** preliminary amendment.
 14. ☐ A substitute specification.
 15. ☐ A change of power of attorney and/or address letter.
 16. ☐ Other items or information:

17. ☒ The following fees are submitted:**Basic National Fee (37 CFR 1.492(a)(1)-(5):**

Search Report has been prepared by the EPO or JPO \$840.00

International preliminary examination fee paid to USPTO (37 CFR 1.482)
..... \$670.00No international preliminary examination fee paid to USPTO (37 CFR 1.482)
but international search fee paid to USPTO (37 CFR 1.445(a)(2)) \$760.00Neither international preliminary examination fee (37 CFR 1.482) nor
international search fee (37 CFR 1.445(a)(2)) paid to USPTO \$970.00International preliminary examination fee paid to USPTO (37 CFR 1.482)
and all claims satisfied provisions of PCT Article 33(2)-(4) \$96.00**ENTER APPROPRIATE BASIC FEE AMOUNT = \$ 970.00**Surcharge of **\$130.00** for furnishing the oath or declaration later than ☐ 20 ☐ 30
months from the earliest claimed priority date (37 CFR 1.492(e))

\$ 0.00

Claims	Number Filed	Number Extra	Rate		
Total Claims	17 -20 =	0	X \$18.00	\$ 0.00	
Independent Claims	3 -3 =	0	X \$78.00	\$ 0.00	
Multiple dependent claim(s) (if applicable)			+ \$260.00	\$ 0.00	

TOTAL OF ABOVE CALCULATIONS = \$ 970.00Reduction by 1/2 for filing by small entity, if applicable. Verified Small Entity statement
must also be filed. (Note 37 CFR 1.9, 1.27, 1.28).

\$ 0.00

SUBTOTAL = \$ 970.00Processing fee of **\$130.00** for furnishing English translation later the ☐ 20 ☐ 30
months from the earliest claimed priority date (37 CFR 1.492(f)).

+ \$ 0.00

TOTAL NATIONAL FEE = \$ 970.00Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be
accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31). \$40.00 per property +

\$ 0.00

TOTAL FEES ENCLOSED = \$ 970.00

Amount to be:	
refunded \$	
charged \$	

- a. ☒ A check in the amount of **\$970.00** to cover the above fees is enclosed.
- b. ☐ Please charge my Deposit Account No. 19-0741 in the amount of \$ to the above fees. A duplicate copy of this sheet is enclosed.
- c. ☒ The Commissioner is hereby authorized to charge any additional fees which may be required, or credit any overpayment to Deposit Account No. 19-0741. A duplicate copy of this sheet is enclosed.

NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.

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29,768

REGISTRATION NUMBER

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IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Attorney Docket No. 016786/0215

In re patent application of
Frederick A.O. MENDELSON et al.
Serial No. Unassigned
Filed: January 8, 1999
For: NEUROACTIVE PEPTIDE

PRELIMINARY AMENDMENT

Assistant Commissioner for Patents
Washington, D.C. 20231

Sir:

Prior to examination of the above-identified application, Applicants respectfully request that the following amendments be entered into the application:

IN THE CLAIMS:

Claim 4, line 1, delete "any one of claims 1 to 3" and insert --claim 1--.

Claim 5, line 1, delete "any one of claims 1 to 4" and insert --claim 1--.

Claim 9, line 1, delete "any one of claims 6 to 8" and insert --claim 6--.

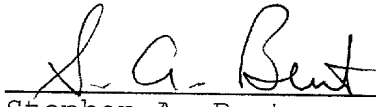
Claim 10, line 1, delete "any one of claims 1 to 9" and insert --claim 1--.

REMARKS

Applicants respectfully request that the foregoing amendments to Claims 4, 5, 9, and 10 be entered in order to avoid this application incurring a surcharge for the presence of one or more multiple dependent claims.

Respectfully submitted,

January 8, 1999



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NEUROACTIVE PEPTIDE

This invention relates to neuroactive peptides, and in particular to peptides which have the ability to act as analogues of angiotensin IV. The peptides of the invention bind with high affinity and specificity to a variety of sites in the central nervous system, and are useful as modulators of motor and cognitive function, and of neuronal development.

Background of the Invention

The renin-angiotensin system has diverse roles in the regulation of body fluid and electrolyte balance and blood pressure control. These actions are exerted in a variety of target organs, including the cardiovascular system, adrenal glands, kidney and central and peripheral nervous systems, by both the circulating hormone and hormone locally produced in tissues. Most of these actions are exerted by the octapeptide, angiotensin II, although the C-terminal heptapeptide angiotensin III has some activity. The hexapeptide NH₂-Val Tyr Ile His Pro Phe-COOH, corresponding to the 3-8 fragment of angiotensin II (ie. amino acids 3-8), is also called angiotensin IV (Ang IV), and has until recently been believed to be an inactive degradation product devoid of biological activity.

However, Harding and co-workers have confirmed an earlier report (Braszko et al, 1988) that Ang IV has central nervous system activity, and can modify learning and behaviour (Wright et al, 1995). In addition, Ang IV has vasoactive effects, and can dilate cerebral arteries (Haberl et al, 1991) and increase renal blood flow (Swanson et al, 1992). This, coupled with the discovery of highly specific, high affinity sites for Ang IV binding in bovine adrenal and other tissues, has reawakened interest in the hexapeptide, and the subject has been comprehensively reviewed (Wright et al 1995).

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Ang IV has been associated with the central nervous system effects of increasing stereotypy behaviour (Braszko et al, 1988) and facilitating memory retrieval in passive avoidance studies (Braszko et al, 1988; Wright et al, 1995). Ang IV also dilates cerebral arterioles (Haberl et al, 1991), and increases renal blood flow (Swanson et al, 1992).

Receptor autoradiographic studies have revealed a widely abundant but selective and characteristic distribution of binding sites for [125 I]Ang IV (known as the AT₄ receptor) in the guinea pig, sheep and monkey central nervous systems, in regions associated with cholinergic neurons and in somatic motor and sensory associated areas (Miller-Wing et al, 1993; Moeller et al, 1995, Moeller et al, 1996). In addition, Ang IV binding sites are abundant in supraspinal components of the autonomic nervous system, and in the spinal cord are found in sympathetic preganglionic neurons, in the dorsal root ganglia, and in Lamina II of the dorsal horn, and in the motor neurons of the ventral horn (Moeller et al, 1995).

The distribution of the Ang IV binding site differs from the localization of the Ang II AT₁ or AT₂ receptors. In addition, the pharmacology of each receptor is distinct in that the Ang IV site exhibits a low to very low affinity for [Sar¹Ile⁸]Ang II, the non-subtype selective Ang II antagonist, and losartan (du Pont-Merck) and PD 123319 (Parke-Davis), the specific AT₁ and AT₂ receptor antagonists respectively (Miller-Wing et al, 1993; Swanson et al, 1992; Hanesworth et al, 1993). Conversely, Ang II receptors show a low affinity for the Ang IV binding site (Bennett and Snyder, 1976).

The wide distribution of the Ang IV binding site in motor, sensory and cholinergic regions suggests important roles for this peptide in the central nervous system. However, a physiological action of the peptide in neurons has yet to be clearly defined.

Numerous neurotransmitters and neuropeptides have

amino acid analogues. Thus the analogue may be a peptidomimetic compound.

Preferably the mammal is a human.

The Ang IV agonist and antagonist compounds
5 according to the invention are useful in the treatment of a variety of conditions, including but not limited to:

- Dementia, including Alzheimer's disease
- Other neurodegenerative disorders involving cholinergic pathways, motor pathways, or sensory pathways,
10 such as motor neurone disease
- sensory and motor peripheral neuropathies
- brain or spinal cord injury due to trauma, hypoxia or vascular disease.

In a second aspect, the invention provides a non-
15 peptide analogue of the peptide of the invention. This non-peptide analogue is to be understood to encompass modifications or substitutions of the peptide structure which are designed to improve the bioavailability, metabolic stability, half-life in the body, or to modify
20 the biological activity, of the compound of the invention. Such non-peptide analogues are known in the art, for example compounds in which the peptide backbone is replaced by a non-peptide chain, and are often referred to as peptidomimetic compounds. Alternatively, in one or more of
25 the peptide linkages the order of the nitrogen and carbon atoms can be reversed to form a pseudo peptide bond. One or more of the amino acid side-chains may be replaced by an analogous structure of greater stability. Many other such variations will occur to the person skilled in the art.
30 The only requirement is that the overall 3-dimensional structure is sufficiently preserved that ability to bind to the AT₄ receptor at suitable affinity is retained. Using modern methods of peptide synthesis and combinatorial chemistry, it is possible to synthesize and test very large
35 numbers of analogues within a short space of time, and such synthesis and screening is routinely carried out by pharmaceutical companies.

Considerable information is available regarding the structural features of Ang IV peptides which are necessary for high affinity, and these results may be used as guidelines for modification of the peptides of the invention. See for example Wright *et al*, 1995.

The person skilled in the art will appreciate that by modifying the sequence or by constructing a non-peptide analogue the activity of the compound of the invention can be very considerably modified. Not only can improvement in activity be obtained, it is also possible to obtain compounds which bind to the AT₄ receptor in such a way that Ang IV activity is inhibited. Such inhibitory compounds can have the ability to antagonize the activity of Ang IV. The person skilled in the art will readily be able to synthesize modified peptides and peptide analogues and to test whether they have activity as Ang IV agonists or antagonists, using methods well known in the art.

According to a third aspect, the invention provides a method of screening for putative agonists or antagonists of the effect of LVV-haemorphin-7 on neuronal activity, comprising the step of testing the ability of the compound to stimulate or inhibit the effect of LVV-haemorphin-7 on a biological activity selected from the group consisting of modifying learning, modifying behaviour, vasoactive effects, dilation of cerebral arteries, increase in renal blood flow, increase in stereotypy behaviour, facilitating memory retrieval, neurite modelling and alleviation of the effects of spinal cord injury.

Thus according to a fourth aspect, the invention also provides compounds which are able to act as agonists or antagonists of the neuroactive peptides of the invention.

Detailed Description of the Invention

The invention will be now described in detail by way of reference only to the following non-limiting

E13 chicken chorioallantoic membranes with varying concentrations of unlabelled compounds: ▲ Ang IV, ◇ Nle¹-AIV, △ CGP 42112, □ Ang II, ▼ Nle¹-Y-I-amide, ★ WSU-4042, ■ [Sar¹Ile⁸]Ang II, ● PD 123319 and

- 5 ○ losartan. Values are expressed as a percentage of total binding, and are pooled from two experiments. B/Bo x 100=% of available receptors occupied;

Figure 3 summarizes competition binding studies showing the inhibition of ¹²⁵I[Sar¹Ile⁸]Ang II binding to
10 E13 chicken chorioallantoic membranes with varying concentrations of unlabelled compounds:

- [Sar¹Ile⁸]Ang II, □ Ang II, △ CGP 42112, ◇ Nle¹-AIV, ▲ Ang IV, ○ losartan, ● PD 123319, ★ WSU-4042 and ▼ Nle¹-Y-I-amide. Values are expressed as a percentage of
15 total binding, and are pooled from two experiments. B/Bo x 100 = % of available receptors occupied;

Figure 4 shows the effect of Ang IV on neurite outgrowth from E11 chicken sympathetic neurons. Values are expressed as a percentage of control levels, and are
20 depicted as the mean±standard error of the mean (SEM). The results are pooled from 3 experiments, each with at least 40 neurite measurements. * indicates a significant difference from control values using Bonferroni's test;

Figure 5 shows the effect of 10 nM Ang IV on
25 neurite outgrowth in the presence of 1 µM Nle¹-Y-I-amide, WSU-4042, Nle¹-AIV, [Sar¹Ile⁸]Ang II, losartan, PD 123319 and CGP 42112. Values are expressed as a percentage of control levels, and are depicted as the mean±S.E.M. The results are pooled from 3 experiments, each with at least
30 40 neurite measurements. * indicates a significant difference from control values using Bonferroni's test;

Figure 6 shows the effect of 10 nM Ang II on neurite outgrowth in the presence of 1 µM Nle¹-Y-I-amide, WSU-4042, Nle¹-AIV, [Sar¹Ile⁸]Ang II, losartan, PD 123319
35 and CGP 42112. Values are expressed as a percentage of control levels, and are depicted as the mean±S.E.M. The results are pooled from 3 experiments, each with at least

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40 neurite measurements. * indicates a significant difference from control values using Bonferroni's test;

Figure 7 illustrates the binding of ^{125}I -angiotensin IV to sheep spinal cord. The arrow indicates the site of damage to the spinal cord;

Figure 8 summarizes the results of competition binding studies showing the inhibition of [^{125}I]LVV-haemorphin-7 binding to sheep cerebellar cortical membranes with varying concentrations of unlabelled compounds:

▲ Ang IV, △ LVV-haemorphin-7, ■ Ang III, □ Ang II, ○ PD 123319, ● losartan, * naloxone and ▽ haloperidol. Values are the mean of three experiments. $B/B_0 \times 100 = \%$ of available receptors occupied;

Figure 9 summarizes the results of competition binding studies showing the inhibition of [^{125}I]Ang IV binding to sheep cerebellar cortical membranes with varying concentrations of unlabelled compounds: ▲ Ang IV, △ LVV-haemorphin-7, ■ AngIII, □ AngII, ○ PD 123319, ● losartan, * naloxone and ▽ haloperidol. Values are the mean of three experiments. $B/B_0 \times 100 = \%$ of available receptors occupied;

Figure 10 is a schematic diagram illustrating the position of the oligonucleotide probes used for cloning and PCR experiments. (A) schematic diagram of the β -globin precursor showing relevant position and direction of oligonucleotides used. The shaded region represents the LVV-haemorphin-7 sequence, which is given below. (B) sequences of the oligonucleotides H170 to H173 (SEQ ID Nos: 2 to 5 respectively) used in this study;

Figure 11 illustrates the detection of β -globin mRNA by RT-PCR and Southern blotting in sheep cerebellar and cerebral cortices, heart and liver. Molecular weight markers are shown on the left;

Figure 12 shows the complete nucleotide sequence of Clone EX (SEQ ID NO:6); and

Figure 13 shows the nucleotide sequence (SEQ ID NO:7) and derived amino acid sequence of the rat EX clone.

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The region of the potential LVV-haemorphin-7 is shown in bold.

Figure 14 summarizes the effects of LVV-haemorphin-7 on the performance of scopolamine-treated rats in a passive avoidance task.

Figure 15 summarizes the effects of LVV-haemorphin-7 on the performance of scopolamine-treated rats in a water maze acquisition trial.

The unlabelled ligands, Ang IV (Peninsula Laboratories, California USA), Ang II and the Ang II antagonist [Sar¹Ile⁸]Ang II (Sigma, Missouri USA), the Ang II partial agonist CGP 42112 (Ciba-Geigy, Basle Switzerland), the Ang II AT₁ antagonist, losartan (Du Pont Merck Pharmaceutical Company, Delaware USA), the Ang II AT₂ antagonist, PD 123319 (Parke-Davis, Michigan USA-Ms. C.L.Germain), and the Ang IV analogues, WSU 4042, Nle¹-Y-I-amide and Nle¹-AIV (prepared as previously described by Sardinia et al, 1993), were used at final concentrations ranging from 10⁻⁹ to 10⁻⁴ M.

Example 1 Mapping of Angiotensin AT₁ Receptors in Monkey Brain

We mapped the distribution of the receptors for Ang IV (AT₁ receptors) in the *Macaca fascicularis* brain using in vitro receptor autoradiography in order to determine if the widespread and distinct distribution of the receptors that are found in the guinea pig brain is also found in primates. The binding sites were initially characterized pharmacologically in competition studies on prefrontal cortical brain sections. These results are summarized in Figure 1. Ang IV, Ang III and Ang II competed for [¹²⁵I]Ang IV binding with IC₅₀s of 5 nM, 80 nM and 730 nM respectively, while Ang II(1-7) was a weak competitor (IC₅₀ of 24 mM). The AT₁ receptor antagonist, losartan (du Pont-Merck) and the AT₂ receptor antagonist, PD 123319 (Parke-Davis), were inactive, even at concentrations of 10 mM. These pharmacological properties

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are similar to those previously described for the AT₄ receptor in bovine adrenal and guinea pig septal membranes, confirming that we were mapping the distribution of the same receptor.

- 5 The distribution of the AT₄ receptor was remarkable, in that its distribution extended throughout several neural systems. This is summarized in Table 1. The most striking finding was the localization of this receptor in motor nuclei and motor-associated regions.
- 10 These included the ventral horn spinal motor neurons, all cranial nerve motor nuclei including the oculomotor, trochlear, facial and hypoglossal nuclei, and the dorsal motor nucleus of the vagus. Receptors were also present in the vestibular, reticular and inferior olivary nuclei, the
- 15 granular layer of the cerebellum, and the Betz cells of the motor cortex. Moderate AT₄ receptor density was seen in all cerebellar nuclei, ventral thalamic nuclei and the substantia nigra pars compacta, with a lower receptor density being observed in the caudate nucleus and putamen.
- 20 The localization of the AT₄ receptor in all levels of the motor hierarchy in the central nervous system implies an important role for the binding site in motor activity.

Accepted for publication

Table 1

Localization and Quantitation of the
AT₄ Receptor in the *Macaca fascicularis* Brain

5

Region	AT ₄ receptor density dpm/mm ² (mean \pm SD)
Caudate nucleus	48 \pm 2
Vertical limb of the diagonal band*	86 \pm 3
Basal nucleus of Meynert*	81 \pm 5
Granular layer of the dentate gyrus	117 \pm 11
CA1	45 \pm 4
CA3	41 \pm 3
Supraoptic retrochiasmatic nucleus*	93 \pm 7
Ventral posterior lateral/medial nuclei	35 \pm 2
Red nucleus*	44 \pm 2
Oculomotor nucleus*	44 \pm 1
Pontine nuclei	50 \pm 2
Lateral geniculate	52 \pm 2
Mo5*	84 \pm 3
Facial nucleus*	90 \pm 4
Hypoglossal nucleus*	93 \pm 8
Inferior olive	76 \pm 10
Granular layer of the cerebellum	126 \pm 10
Molecular layer of the cerebellum	47 \pm 6

Values are the mean of four sections from one animal and are representative of the relative densities of AT₄ receptors. * Values are determined from the overall area and not from individual cell bodies which exhibit higher binding.

In addition to the somatic motor nuclei and autonomic preganglionic motor nuclei, abundant AT₄ receptors were also found in other cholinergic systems and

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their projections, including the nucleus basalis of Meynert, vertical limb of the diagonal band and the hippocampus. Apart from being a neurotransmitter in motor neurons, acetylcholine is also implicated in cognition, since anti-cholinergic drugs induce memory disorders and confusion; in Alzheimers's disease, neuronal loss occurs in the cholinergic-rich basal nucleus of Meynert. Ang IV has been shown by two independent studies to facilitate memory retrieval in passive and conditioned avoidance tests (Braszko et al, 1988; Wright et al, 1993), and, when administered intracerebroventricularly, induces c-fos expression in the hippocampus (Roberts et al, 1995). Together with the presence of high densities of AT₄ receptors in this region, these observations suggest that Ang IV may play an important role in the modulation of cognitive function.

AT₄ receptors were also observed in sensory regions, with moderate levels in spinal trigeminal, gracile, cuneate and thalamic ventral posterior nuclei, and in the somatosensory cortex. While receptor density was low in sensory neurons when compared with that observed in motor and cognitive areas, the AT₄ receptor was located throughout most sensory-associated areas, including the lamina II of the spinal cord, gracile, cuneate and spinal trigeminal nuclei, ventral posterior thalamic and lateral geniculate nuclei and the sensory cortex, suggesting a substantial involvement with sensory activity. This distribution pattern has also been observed in the guinea pig and sheep brain. As shown in Example 2, abundant AT₄ receptors were also observed in sheep dorsal root ganglia.

Example 2 **Mapping of Angiotensin AT₄ Receptors in**
Sheep Spinal Cord

We extended the localization of the AT₄ receptors to the sheep spinal cord, to investigate if the strong presence of the AT₄ receptors in supraspinal motor and sensory regions persists in the spinal cord.

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When the binding characteristics of [125 I]Ang IV were assessed in the eighth cervical segment (C8) of the sheep spinal cord, we found that the affinities of the different unlabelled ligand in competing for the binding were similar to those observed for the monkey brain.

In the sheep spinal cord, high densities of AT₄ receptors were found in lamina IX in the ventral horns of all segments examined. At a cellular level, the binding was found overlying the cytoplasm of lateral and medial motor neurons and in their processes, but binding was absent from the cell nuclei. Whilst a clearly defined function of the Ang IV binding site is yet to be determined, the association with motor activity is strengthened in view of its abundant localization in the motor neurones in the ventral horn of the spinal cord, in addition to its strong presence in supraspinal motor areas.

High densities of AT₄ receptors were also found in the lateral tip of lamina VII of all thoracic segments and lumbar segments L1 to L4, which corresponded with sympathetic preganglionic neurons in the intermediolateral cell column. However, binding was absent from L5 and L6 and from the sacral segments S1 and S2.

In the dorsal root ganglia associated with all spinal segments, high densities of AT₄ receptors were found in the cytoplasm of small and large cell bodies of the sensory neurons, but not in the satellite cells, nor in the endoganglionic connective tissue. In laminae I and II, the terminal fields of the dorsal root ganglia sensory afferents, only a low abundance of the receptor was noted in lamina II. Despite the low levels of AT₄ receptors in lamina II, their high abundance in the dorsal root ganglia and their consistent but low levels in most supraspinal sensory areas suggest that AT₄ receptors may still play a role in the processing of sensory information.

Low levels of the AT₄ receptors were also found in the blood vessels which extended radially to the pial surface, in the blood vessels of the anterior and posterior

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fissures, and in the ependyma of the central canal. Ang IV has been reported to induce an endothelium-dependent dilation of rabbit pial arterioles, and in rats Ang IV reverses acute cerebral blood flow reduction after experimental subarachnoid haemorrhage.

Our localization studies suggest that AT₄ receptors are quite distinct from the known angiotensin receptors -the AT_{1a}, AT_{1b} and AT₂ receptors - in terms of their pharmacological specificity and their pattern of distribution in the brain and spinal cord. Furthermore, the pattern of distribution of the AT₄ receptors suggests that they may be involved in the function of neurones involved in motor function, sensory function and cholinergic systems, including cognition.

Example 3 Characterization of Embryonic Chicken Ang IV and Ang II Binding Sites

In order to characterize the pharmacology of the embryonic chicken AT₄ and Ang II receptors, chorioallantoic membranes (CAM) from embryonic day 13 (E13) chickens were used. The membranes were removed and frozen in isopentane cooled to -40°C.

a) Characterization of the embryonic chicken Ang IV binding site

CAM were homogenized in 30 ml of a hypotonic buffer (50 mM Tris, pH 7.4, 5 mM EDTA) and then centrifuged for 10 min at 500 g and 4°C. The supernatant fraction was removed and centrifuged for 20 min at 40,000 g and 4°C. The resulting pellet was rehomogenized in 2 ml of hypotonic buffer, and the final volume of the homogenate was adjusted to give a protein concentration of 10 mg/ml, as determined by the Biorad protein assay. The binding assay contained CAM (100 µg of protein), 0.14 µCi of [¹²⁵I]Ang IV (approximately 260 pM), and competing ligand, in a total volume of 270 µl in a 50 mM Tris buffer, pH 7.4, containing 150 mM NaCl, 5 mM EDTA, 100 µM phenylmethylsulfonyl

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fluoride, 20 μ M bestatin and 0.1% (w/v) bovine serum albumin. The binding system was incubated at 37°C for 2 h.

5 b) Characterization of the embryonic chicken Ang II binding site

CAM were prepared as described above with the following exceptions. The isotonic buffer contained 50 mM Tris, pH 7.4 and 6.5 mM $MgCl_2$ and the hypotonic buffer contained 50 mM Tris, pH 7.4, 6.5 mM $MgCl_2$, 125 mM NaCl and 10 0.2% (w/v) bovine serum albumin. In addition, the peptidase inhibitors, leupeptin, lisinopril, phosphoramidon, Plummer's inhibitor and bestatin, each used at a 1 μ M concentration and 1 mM benzamidine and 2.5 mM phenanthroline, were included in both buffers.

15 In binding competition studies on E13 chicken CAM, [^{125}I]Ang IV binding was strongly inhibited by Ang IV and Nle¹-AIV (IC₅₀s of 18 and 43 nM respectively), whereas WSU-4042, Nle¹-Y-I-amide and Ang II were weaker competitors with IC₅₀s of 5, 2.2 and 0.65 μ M respectively, and losartan 20 and PD 123319, were inactive at concentrations up to 10 μ M. [Sar¹Ile⁸]Ang II and CGP 42112 were effective at only competing for 50% of the sites, and then only at concentrations of 10 and 0.5 μ M respectively. These results are summarized in Figure 2.

25 In studies of [^{125}I][Sar¹Ile⁸]Ang II binding to CAM, Ang II, [Sar¹Ile⁸]Ang II and CGP 42112 competed for binding with IC₅₀s of 100, 13 and 180 nM respectively, whilst Ang IV, Nle¹-AIV and losartan were very weak competitors (IC₅₀s of 50, 8 and 100 μ M respectively). PD 123319, 30 WSU-4042 and Nle¹-Y-I-amide exhibited IC₅₀s greater than 100 μ M. These results are shown in Figure 3.

Example 4 Effects of Ang IV on Neurite Outgrowth

35 The wide distribution of the AT₄ receptors in motor, sensory and cholinergic regions suggests important roles for this peptide in the central nervous system. However, a physiological action of Ang IV in neurons has

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yet to be clearly defined. Numerous neurotransmitters and neuropeptides have been associated with the regulation of neuronal development. For instance, acetylcholine inhibits neurite outgrowth from embryonic chicken ciliary ganglion cells, sympathetic neurons, and rat hippocampal neurons. Conversely, vasoactive intestinal peptide stimulates superior cervical ganglion branching and somatostatin increases neuronal sprouting from *Helisoma* buccal ganglion neurons.

We determined whether Ang IV has a trophic role in the central nervous system by examining its effects on neurite outgrowth from cultured embryonic chicken sympathetic neurons.

Sympathetic ganglia from E11 chickens were dissociated using trypsin/Versene, and were cultured in 24 well plates in DMEM and Ham's F12 medium which contained 1% (v/v) insulin-transferrin-selenium-X growth supplement (Gibco BRL, Maryland USA), 100 mM putrescine, 1.67 mg/ml prostaglandin F2 α , 6.67 ng/ml progesterone, and 5 ng/ml nerve growth factor (Sigma, Missouri USA). Neurons were allowed to adhere to the wells (approximately 2 h) before being given a 24 h treatment of peptides and/or their antagonists. Peptides and antagonists used were added to the cultures 0.5 h prior to either Ang IV or Ang II addition. Ang IV dose response curves were performed over the concentration range 10⁻¹¹ to 10⁻⁵ M. Culture dishes were coated with 0.1 mg/ml polylysine and then given three washes with phosphate-buffered saline (PBS) before being coated with 10 μ g/ml laminin. Wells were washed with PBS before being used for culture.

At the conclusion of the experiment, the culture medium was removed from the wells, the neurons were fixed with 2.5% glutaraldehyde in PBS for 20 min and examined under a phase-contrast microscope, attached to an MD30 Plus image analysis software (Adelaide, Australia). The length of neurites (longer than 50 μ m) of every neuron examined was measured. A minimum of forty neurite measurements was

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taken per treatment group, and each experimental treatment was performed at least in triplicate.

At the conclusion of the experiment, the viability of the cells were confirmed by exclusion of 0.1% aniline blue.

In cultures of embryonic (E11) chicken sympathetic neurons, Ang IV inhibited neurite outgrowth in a dose-dependent manner, with a threshold at 10^{-11} M, half maximal inhibition at 10^{-10} M and a maximal effect at 10^{-9} M. Between 10^{-9} to 10^{-5} M, outgrowth was maximally inhibited ($P < 0.05$). These results are shown in Figure 4. At 10^{-8} M Ang IV, the inhibition of neurite outgrowth was totally reversed by 1 μ M of the Ang IV analogues WSU-4042, Nle¹-Y-I-amide, and Nle¹-AIV. The effects of the analogues alone were not statistically different from control values. In contrast to the Ang IV analogues, the Ang II antagonist, [Sar¹Ile⁸]Ang II, the AT₁ and AT₂ antagonists, losartan and PD 123319, and the Ang II partial agonist, CGP 42112, had no effect on the Ang IV response, as shown in Figure 5.

At 10^{-8} M Ang II, neurite outgrowth was inhibited by 25%, which was highly significant. The Ang IV analogues completely reversed this effect, whilst the Ang II antagonists [Sar¹Ile⁸]Ang II, losartan, PD 123319, and CGP 42112 were ineffective. This is illustrated in Figure 6.

These studies suggest that the inhibition of neurite outgrowth by both peptides is mediated by the AT₄ receptors, and supports a role for angiotensin IV in neurite modelling.

Example 5 Effect of Angiotensin IV on Spinal Cord Damage

Glial fibrillary acid protein (GFAP)-positive astrocytes are involved with modelling neurite formation after damage to the spinal cord (Bovolenta et al, 1992). Injury-evoked plasticity is a similar situation to that observed in the developing embryo (Schwartz, 1992). In

light of our findings on the ability of spinal cord tissue to bind Ang IV (Example 2), we tested the effect of spinal cord injury on Ang IV binding. Surprisingly, we found a marked elevation of [125 I]Ang IV binding in damaged spinal cord sections. This is illustrated in Figure 7.

These results suggest that the AT₄ receptor may be a suitable target for alleviation of the effects of spinal cord injury.

10 Example 6 Purification of an Endogenous Brain Peptide
 Which Binds to the AT₁ Receptor

The level of Ang IV in the brain is very low to undetectable (DJ Campbell, personal communication). The widespread and characteristic distribution of AT₄ receptors in the central nervous system suggests that there may be an as yet unidentified peptide ligand for this receptor. We therefore undertook a search for such a ligand, using conventional protein chemistry purification techniques together with an AT₄ receptor assay system in order to detect and monitor substance(s) in extracts of sheep brain which compete for [¹²⁵I]Ang IV binding in this system.

a) $^{125}\text{AT}_4$ Receptor Binding Assay

The binding of ^{125}I -Ang IV to bovine adrenal membranes was used as an assay system to screen for AT_4 receptor binding activity in sheep cerebral cortex fractions. Bovine adrenal glands obtained from the abattoir were diced into 1 mm x 1 mm blocks, homogenized in 3 ml of a hypotonic buffer (50 mM Tris, 5 mM EDTA, pH 7.4) and then centrifuged for 10 min at 500 g. The supernatant was removed and centrifuged for 20 min at 40,000 g, and the resulting pellet was rehomogenized in 2 ml of hypotonic buffer. Binding assay samples contained bovine adrenal (56 mg of protein as determined by the Biorad protein assay), 0.14 μCi of ^{125}I -Ang IV (approximately 260 pM), and 10 μl of test sample, in a total volume of 270 μl in 50 mM Tris buffer, pH 7.4, containing 150 mM sodium

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chloride, 5 mM EDTA, 100 μ M phenylmethanesulfonyl fluoride, 20 μ M bestatin, and 0.1% (w/v) bovine serum albumin. The relative potency of the fractions in competing for 125 I-Ang IV binding was determined from a standard curve in which known amounts of unlabelled Ang IV were added (10^{-10} to 10^{-6} M). Fractions from each purification step were assayed for their ability to compete for [125 I]Ang IV binding, with those exhibiting the highest activity undergoing the next purification step.

10

b) Purification Procedure

Sheep cerebral cortex was homogenized in 2 M acetic acid, (2 ml/g tissue), centrifuged, and the supernatant decanted. A preliminary purification of the extract was performed using a column of preparative C18 material (55-105 mm, Waters). The C18 eluent was lyophilized, reconstituted, and subjected to a series of chromatographic steps, in which fractions were assayed for Ang IV displacement activity. In brief, the chromatographic steps were: three successive reversed-phase HPLC steps, using columns of varying pore size (Deltapak C18, 300 $^{\circ}$ A, and Novapak C18) as well as changing ion-pairing agents, solvents and gradient elution conditions; this was followed by anion exchange, then cation exchange, with final purification on a microbore LC C8 column. The purified active peptide was sequenced using an Applied Biosystems Model 470 A Protein Sequencer with an on-line Model 120A PTH Analyzer.

The sheep cerebral cortex yielded 1.9 nmoles of AT₄ receptor binding activity per gram of wet weight after the first C18 Deltapak column. Following the third Poly LC column (55 $^{\circ}$ C), Ang IV activity coeluted with the major UV absorption peak, and the following peptide sequence was obtained from this peak:

Leu-Val-Val-Tyr-Pro-Trp-Thr-Gln-Arg-Phe (SEQ ID NO:1).

A search of protein database records revealed that this sequence corresponded to the amino acid sequence

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32-41 of the human β , δ , γ and ϵ globin chains and is known as LVV-haemorphin-7.

LVV-haemorphin-7 is a 10 amino acid peptide found in the brain, pituitary, hypothalamus and bone marrow which binds with high affinity to the angiotensin AT₄ receptor. The sheep peptide sequence is identical to amino acids 30-39 of the sheep β_A , β_B , β_C , and ϵ globin precursors (Garner and Lingrel, 1989; Saban and King, 1994), and this sequence is conserved in many species, including human (see for example Karelin *et al*, 1994). In humans, there are 6 β -globin-like genes ϵ , γ^A , γ^F , δ , β and a pseudogene $\psi\beta$, clustered on chromosome 11, and all encode the LVV-haemorphin-7 sequence (Karlsson and Nienhuis, 1985). This sequence is not present in any of the α globin family of genes. LVV-haemorphin-7 and some shorter sequences within this peptide have opioid activity, and it appears that the sequence VVYP is required for this activity (Karelin *et al*, 1994).

20 Example 7 Properties of Synthetic LVV-haemorphin-7

A decapeptide with the sequence isolated above was synthesized under contract by Chiron Mimotopes, and its biochemical and pharmacological properties were characterized as follows:

25

a) HPLC

A preliminary high performance liquid chromatography (HPLC) run indicated that the synthetic peptide did not coelute with the fraction that was sequenced. It appeared that the fraction might have been degraded due to prolonged storage at 4°C. Mass spectrometry analysis was carried out in order to determine whether this was the case. The data obtained from mass spectrometry analysis of the two active peaks produced following prolonged storage of the original purified material were indeed consistent with degradation. The early eluting peak gave a mass corresponding exactly to the loss

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of the phenylalanine residue from the carboxy terminus, whereas the second active peak gave a mass corresponding exactly to the loss of the amino terminal leucine residue. Furthermore, these data (given that all the mass readings were unambiguous) strongly suggest that the active peptide is not post-translationally modified, either in the peptide core or at the amino or carboxyl terminus.

b) Ligand Binding Studies

The pharmacological properties of the decapeptide LVV-haemorphin-7 in competing for the binding of ^{125}I -Ang IV in bovine adrenal membrane and sheep cerebellar cortical membranes were determined. Both LVV-haemorphin-7 and Ang IV were radioiodinated using chloramine T, and separated on a C18 Sep-pak column using 0.5% trifluoroacetic acid in a 20-80% methanol gradient.

Bovine adrenal membranes or sheep cerebellar cortical membranes were homogenized in 30 ml of a hypotonic buffer (50 mM Tris, 5 mM EDTA, pH 7.4), and then centrifuged for 10 min at 500 g. The supernatant was removed and centrifuged for 20 min at 40,000 g, and the resulting pellet was rehomogenized in 2 ml of hypotonic buffer. Binding assays contained:

bovine adrenal (56 μg of protein) or sheep cerebellar membranes (26 μg of protein), as determined by the Biorad protein assay (Bradford, 1976);

0.14 μCi of [^{125}I]Ang IV (approximately 260 pM), or 0.11 μCi of [^{125}I]LVV-haemorphin-7 (approximately 200 pM), and

competing ligand,

in a total volume of 270 μl in 50 mM Tris buffer, pH 7.4, containing 150 mM sodium chloride, 5 mM EDTA, 100 μM phenylmethylsulfonyl fluoride, 20 μM bestatin and 0.1% (w/v) bovine serum albumin.

The assay was incubated at 37°C for 2 h.

In the bovine adrenal membranes, a range of concentrations of unlabelled LVV-haemorphin-7 or Ang IV was

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added to the assay system in order to determine the relative potencies of the two peptides in this radioreceptor assay system. Both Ang IV and LVV-haemorphin-7 displayed comparable affinities in competing for the ^{125}I -Ang IV binding (approx. 1-5 nM), with Ang IV exhibiting slightly higher affinity.

For competition studies in sheep cerebellar cortical membranes, dilutions of the unlabelled ligands, LVV-haemorphin-7, Ang IV, Ang II, Ang III and the non-specific opioid antagonist, naloxone, the Ang II AT₁ antagonist, losartan, the Ang II AT₂ antagonist, PD 123319, and the sigma opioid and dopamine D₂ antagonist, haloperidol, were used at concentrations ranging from 10^{-13} to 10^{-4} M. Quantitation of receptor binding was calculated as the mean of two experiments.

In these studies, ^{125}I -LVV-haemorphin-7 binding to sheep cerebellar cortical membranes was competed for by LVV-haemorphin-7, Ang IV, Ang III, and Ang II (IC₅₀s of 5.6 nM, 1 nM, 77 nM, and 1.6 μM respectively). PD 123319 was a weak competitor (IC₅₀ of 46 μM), whilst losartan, naloxone and haloperidol were ineffective (IC₅₀ greater than 100 nM). These results are illustrated in Figure 8. Similarly, [^{125}I]Ang IV binding to cerebellar membranes was competed for by Ang IV, LVV-haemorphin-7, Ang III, and Ang II with IC₅₀s of 1.13 nM, 2 nM, 6.9 nM and 2 μM respectively, whilst PD 123319, losartan, naloxone and haloperidol were inactive at 10 μM . These results are illustrated in Figure 9.

c) Binding of ^{125}I -LVV-haemorphin-7 to Sheep Brain

Sheep hindbrain sections were used to compare the distribution of ^{125}I -LVV-haemorphin-7 binding and AT₄ receptor sites. Sections at 10 μm thickness were equilibrated to 22°C (30 min), and then preincubated for 30 min in an isotonic buffer containing 50 mM Tris, 150 mM sodium chloride, 5 mM EDTA, 100 μM phenylmethylsulfonyl fluoride, 20 μM bestatin and 0.1% bovine serum albumin,

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pH 7.4, before a further 2 h incubation in the same buffer containing 2.84 μCi of [^{125}I]LVV-haemorphin-7 or [^{125}I]Ang IV (approximately 140 pM). The binding of the radioligands was cross-displaced with either 1 μM unlabelled LVV-haemorphin-7 or Ang IV. After incubation, the sections were given three 2 min washes in buffer at 4°C, and exposed to X-ray film for 14 to 28 d.

[^{125}I]LVV-haemorphin-7 and [^{125}I]Ang IV exhibited an identical binding pattern in the sheep hindbrain. Binding was localized to the motor-associated areas, the granular layer of the cerebellum, the inferior olive, hypoglossal and lateral reticular nuclei, to the autonomic regions, the dorsal motor nucleus of the vagus and the nucleus ambiguus, and to the sensory regions, the external cuneate and spinal trigeminal nuclei. The binding of both radioligands was displaced by a 1 μM concentration of either unlabelled Ang IV or LVV-haemorphin-7, indicating that not only are the two binding sites distributed in the same brain regions, but that the two radioligands are actually binding to the same sites.

Example 8 **Isolation of Potential LVV-Haemorphin-7 Precursor Clones**

It is not known whether LVV-haemorphin-7 is synthesized in the brain, or whether it is derived from the breakdown of haemoglobin. Demonstration of LVV-haemorphin-7 precursor mRNA in the brain would provide evidence for the former. Possible methods to demonstrate that LVV-haemorphin-7 precursor mRNA is present in the brain include:

- (a) isolation of specific cDNA clones from a brain cDNA library;
- (b) detection of the mRNA in the brain by RT-PCR;
- (c) detection of LVV-haemorphin-7 precursor mRNA by *in situ* hybridization histochemistry; and
- (d) demonstration of the mRNA in brain specific

cell cultures.

It has previously been reported that α - and β - globin mRNAs are expressed in mouse brain, as demonstrated by Northern analysis (Ohyagi, Y., et al, 1994).

5 Each of these approaches has specific advantages. *In situ* hybridization histochemistry and detection of the mRNA in brain specific cell cultures would provide evidence for synthesis in the brain. Isolation of clones and the reverse transcription polymerase chain reaction (RT-PCR)
10 detection of mRNA would show the presence of mRNA in the brain, but contamination by reticulocytes cannot be excluded. However, isolation of cDNA clones provides considerable information about the structure of the precursor. The precursor of LVV-haemorphin-7 may be a
15 member of the β -globin family, eg β^A etc, or an alternatively spliced globin, or it may be a previously unknown non-globin peptide.

To isolate potential clones that code for the precursor of the LVV-haemorphin-7 peptide, we have screened
20 a rat brain cDNA library using an oligonucleotide based on the LVV-haemorphin-7 sequence.

Oligonucleotide Design

A number of oligonucleotides have been designed,
25 as illustrated in Figure 10. Oligonucleotide H170 (SEQ ID NO:2) was designed to correspond to the region of the sheep β -globin gene encoding the LVV-haemorphin-7 sequence. This probe was used for screening the library, and also as the sense oligonucleotide in PCR. Oligonucleotide H173 (SEQ ID
30 NO:5) was designed as the antisense primer for use in PCR. PCR with H170/H173 spans intron 2, and will generate a 255 bp fragment with cDNA as the template and a 1098 bp fragment with genomic DNA. Oligonucleotide H172 (SEQ ID NO:4) can be used as an internal probe for H170/H173 PCR
35 products. Oligonucleotide H172 and H173 (SEQ ID NO:4, 5) are antisense probe corresponding to exon 2 and 3 of the sheep β -globin gene, and were used for *in situ*

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hybridization histochemistry.

Detection of β -Globin Like Sequences in Brain by Polymerase Chain Reaction (PCR)

5 RNA was isolated from sheep cerebellar and cerebral cortices, heart and liver. The RNA (20 μ g) was reverse transcribed in a 25 μ l reaction containing 100mM KCl, 50mM Tris-HCl (pH 8.4), 6 mM $MgCl_2$, 10 mM dithiothreitol, 500 μ M dNTPs (Progen), 12 μ g/ml random
10 hexamers (Boehringer Mannheim), 40 units RNasin (Progen), and AMV reverse transcriptase (Boehringer Mannheim, 25 units) at 42°C for 1 h. An aliquot of the reverse transcription reaction (10%) was used in the polymerase chain reaction. The primers used for amplification of the
15 β -globin mRNA were sense H170 and antisense H173 (see Figure 10). PCR was performed in a reaction containing: 10mM Tris-HCl (pH 8.3), 50mM KCl, 400 μ M dNTPs, Taq Polymerase (Bresatec, 2.5 units), 3 μ M $MgCl_2$, and each primer at 400nM. Denaturation, annealing and extension were
20 carried out at 94°C, 60°C and 72°C for 1 min each for 40 cycles, followed by a final extension at 72°C for 10 min.

The PCR products were separated on an agarose gel, transferred to Hybond N+, and Southern analysis using an internal oligonucleotide (H172) was performed to confirm
25 that the products were derived from globin precursors. Specific bands of the expected size of 255 bp were detected in all four tissues examined, as shown in Figure 11.

Screening a Rat cDNA Library for β -Globin Like Sequences

30 An oligonucleotide corresponding to the nucleotide sequence of the LVV-haemorphin-7 region of the sheep β -globin (H170) was used to screen a rat brain cDNA library (Stratagene Cat No: 936515, Sprague-Dawley, whole brain). Approximately 8×10^5 clones were plated, and
35 plaque lifts taken using standard methods (eg Maniatis et al: Molecular Cloning). The filters were prehybridized in Rapid-Hyb (Amersham) for 1 hr at 42°C, then the 5' end

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labelled H170 was added for 2 hr. The filters were then washed 3 times at 42°C in 2xSSC/0.1% SDS. The filters were autoradiographed for 4 days using Biomax film and an intensifying screen. A total of 24 putative positives was isolated. The positives were eluted in PSB.

The positives were then further characterized using a PCR based method. PCR was performed using oligonucleotide H170 as the 5' primer and H173 as the 3' primer. A PCR product derived from H170/H173 will span an intron in the sheep β -globin gene, and will generate a 1098 bp fragment.

An aliquot of the eluted λ clone was boiled for 5 min, then chilled on ice. This was used as template DNA in a PCR reaction containing 10 mM Tris-HCl (pH 8.3), 50 mM KCl, 400 μ M dNTPs, Taq Polymerase (Bresatec, 2.5 units), 3 μ M MgCl₂, and each primer at 400 nM. Denaturation, annealing and extension were carried out 94°C, 60°C and 72°C for 1 min each for 30 cycles, followed by a final extension at 72°C for 10 min. PCR products were analysed by electrophoresis on a 1.4% agarose gel.

The H170 positive/PCR negative clones were stored for further characterization. It is considered that they may be either non-globin precursors, alternatively spliced precursors or fragments of globin clones.

Sequencing Rat β -globin Clones

The 6 positives selected by PCR were plaque purified, and subjected to plasmid excision according to the manufacturers instructions. The insert sizes were determined by separate restriction mapping with the enzymes EcoRI and PvuII. Clones EX, FX, LX, RX and TX contain inserts of approx 500 bp. Clone DX was the longest, and contained an insert of approximately 2500bp. Southern analysis of the clones using an internal oligonucleotide (H172) confirmed that these clones were derived from globin precursors.

These plasmids were sequenced using the Pharmacia

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T7 sequencing kit. Sequencing of clones EX, FX and LX, using the universal primer, showed sequence homology to the 3' untranslated region of β -globin. Clones RX and TX when sequenced with the universal primer, and clone DX when
 5 sequenced with the reverse primer, showed sequence homology to the 5' end of the β -globin gene, including the initiation codon ATG.

Clone DX was subjected to nested deletion analysis to generate more templates for sequencing. This
 10 clone contained the β -globin sequence, and approximately 1.8 kb of sequence which was not homologous to the globin cluster, and may be the result of two inserts in the one clone.

Complete sequencing of clone EX showed that the
 15 clone was identical to rat β^A -globin (Genbank accession No: X16417), as shown in Figure 12. Figure 13 shows the nucleotide sequence and derived amino acid sequence of clone EX, indicating the putative LVV-haemorphin-7 region.

20 Example 9 In situ Hybridization Histochemistry

The distribution of mRNA encoding LVV-haemorphin-7 and its precursor peptide is being investigated using a range of oligonucleotides for the different regions of the β -globin gene, including the C-terminal of exon 2 (H172 of
 25 Figure 13) and the N-terminal regions of exon 3 (H173). The antisense (initially H172, H173) oligonucleotides were 3' end labelled with a ^{35}S -dATP using terminal d-transferase and purified on a Nensorb column. Sheep brain sections were then hybridized with 7.5×10^5 cpm of
 30 labelled probe in a 75 μl total volume of 50% formamide, 4xSSC, 1xDenhardt's solution, 2% sarcosyl, 20 mM Na_2PO_4 buffer (pH 7), 10% dextran sulphate, 50 $\mu\text{g}/\text{ml}$ herring sperm DNA and 0.2 mM dithiothreitol. After a 16 h hybridization period, the sections were washed four times in 1 x SSC,
 35 rinsed in distilled water and dehydrated through increasing ethanol and exposed to Hyperfilm β -max. Preliminary experiments using oligonucleotides H172 and H173 detected

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β -globin mRNA in the inferior colliculus and nucleus of the spinal trigeminal. Further *in situ* hybridization histochemical studies involve the use of additional antisense and sense synthetic oligonucleotides from different regions of the β -globin sequence to confirm our finding of β -globin mRNA in brain nuclei. The distribution of β -globin mRNA is then compared to our autoradiographic localization of the AT₁ receptors in order further to elucidate roles for this novel peptide system.

10

Example 10 Radioimmunoassay and Immunohistochemical
Detection of LVV-haemorphin-7

Two sheep were immunized with the LVV-haemorphin-7 sequence conjugated to diphtheria toxoid and both antisera and affinity purified antisera with adequate titre to set up radioimmunoassays for LVV-haemorphin-7 have been obtained. The radioimmunoassay, which is of conventional type, is used to determine the concentration of LVV-haemorphin-7 in different tissues or in specific regions within a tissue, in order to provide us with further information as to other possible physiological actions of the decapeptide.

The antisera are also used immunohistochemically to determine the tissue distribution of LVV-haemorphin-7, particularly in the brain. Guinea pigs are perfused intracardially with 4% paraformaldehyde in phosphate-buffered saline solution, the tissues dissected out and immersed in a 20% sucrose solution overnight. The tissues are then frozen, 5-10 micron sections cut, and endogenous peroxidase blocked by a 30 min incubation in 0.5% hydrogen peroxidase in methanol prior to an overnight incubation with the primary antibody in phosphate-buffered saline containing 3% normal goat serum. After a few washes in phosphate buffered saline, the sections are incubated with the secondary anti-sheep antibody, and detected using the streptavidin-biotin/horseradish peroxidase complex system (Vectastain). The detection of LVV-haemorphin-7 in neurones

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provides further support that the decapeptide is synthesized within neurones, and thereby may function as a neuropeptide, since we have already shown that its receptor occurs in neurones. Immunohistochemistry is also performed
5 at the electron microscopic level in order to evaluate the subcellular distribution of the peptide, in particular whether it occurs in intracellular storage granules.

The radioimmunoassay for LVV-haemorphin-7 is also employed to investigate the secretion of the peptide from
10 neural tissue. Slices prepared from brain regions found to be rich in LVV-haemorphin-7 immunoreactivity are incubated in Krebs Ringer Bicarbonate buffer at 37°C, and the effects of depolarization by high K⁺ medium and various secretagogues are evaluated to test whether the peptide is
15 secreted from neurones. Similar experiments are carried out on cultured neuronal cell lines which are found to contain the peptide. Radioimmunoassays of body fluids including plasma and cerebrospinal fluid are used to determine levels of the peptide in these fluids under
20 normal and pathological conditions.

In addition, the subcellular distribution of the peptide is evaluated by radioimmunassay of subcellular fractions from nervous tissues, including synaptosomes, in order to evaluate if the peptide is stored in subcellular
25 granules, as occurs for other secreted neuropeptides.

Example 11 Effect of LVV-haemorphin-7 in Passive Avoidance Conditioning Trials

Angiotensin IV has been shown to improve memory
30 retention and retrieval in a passive avoidance task (Braszko et al, 1988, Wright et al, 1993), an effect which was mediated via the AT₁ receptor. Scopolamine, a muscarinic receptor antagonist, has been used to induce amnesia. It has been reported that a more stable analogue
35 of angiotensin IV, WSU 2088, reversed the disruption in learning in a passive avoidance task induced by scopolamine. The effects of LVV-haemorphin-7 on the

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conditioned passive avoidance task in untreated and scopolamine-treated rats were tested.

Rats were surgically implanted with intracerebroventricular cannulae and handled daily. On the conditioning day, each animal was habituated to the dark compartment of a passive avoidance conditioning apparatus for 5 min with the guillotine door closed. The animal was then returned to its home cage for 5 min and then placed in the light compartment with the guillotine door opened.

Latency to enter the dark compartment with all four feet was measured in seconds. These trials were repeated with 5 min in the home cage between trials until the rat entered the dark side within 20 seconds. Before the final trial on conditioning day, the rats were randomly divided into four groups: (a) saline followed by saline (b) saline followed by 1.0 nmol LVV-haemorphin-7 in (c) 70 nmol scopolamine followed by saline (d) 70 nmol scopolamine followed by 1.0 nmol LVV-haemorphin-7, all administered in a volume of 2.5 µl intracerebroventricularly 30 min and 5 min before the final trial respectively. On the last trial, the guillotine door was closed and the animals received one low-level shock (0.2mA) for 1.5 seconds via the grid floor. The animals were then returned to their home cages for 24 hours before being tested once daily for the next four days and the latency periods to reenter the dark compartment were measured. Results are shown in Figure 14.

In this passive avoidance paradigm, the control animals which received successful conditioning displayed high latencies in entering the darkened compartment, whereas rats treated with scopolamine displayed learning and memory deficits, as indicated by much lower latencies in entering the dark compartment. The mean latencies to enter the dark compartment of rats which received LVV-haemorphin-7 after scopolamine were not significantly different from those of the control rats, indicating that in these rats LVV-haemorphin-7 completely reversed the scopolamine-induced amnesia. However, the rats which

- 30 -

received LVV-haemorphin-7 alone performed worse than the scopolamine-treated rats.

These results indicate that LVV-haemorphin-7 successfully counteracts the memory disruption induced by scopolamine treatment. However, administration of the peptide alone was detrimental to learning, which may be due to overstimulation of the neuronal system because of the high dose used.

Effective doses of LVV-haemorphin-7 are determined by conducting dose-response studies with LVV-haemorphin-7 and observing the effects on learning a passive avoidance task in the animals, including those with scopolamine-induced amnesia. Similar studies are also used to determine if the memory disruption caused by LVV-haemorphin-7 is due to excessively high doses of the peptide.

Example 12 **Effect of LVV-haemorphin-7 in the Water Maze Acquisition Trials**

The circular water maze (Morris water maze) consists of a circular tank containing water which has been rendered opaque, with a hidden platform underneath the surface of the water. Scopolamine blocks the trial-to-trial decrease in latency of this task, and this effect appears to be due to impairment of short-term memory. The effect of LVV-haemorphin-7 on the scopolamine-induced amnesia in this task was investigated.

Rats were surgically implanted with intracerebroventricular cannulae and handled daily. On the day of the trial, the rats were introduced into the water maze from different starting positions equidistant from the escape platform. The time taken for each rat to reach the platform was noted. There were four consecutive trials for each animal on each day, with a 60 second rest period between trials. The mean latency period before the animal reached the platform was plotted, and is shown in Figure 15. On days 1 and 2 of the trial (non-spatial), none of

- 31 -

the animals received any drug treatment. Although the scopolamine group displayed increased latency on day 1, the latency on day 2 decreased to control level. The rats were then randomly divided into 3 groups:

- 5 (a) the saline control,
- (b) 70 nmol scopolamine in 2.5 μ l, and
- (c) 70 nmol scopolamine followed by 1.0 nmol LVV-haemorphin-7, and were subjected to 5 days of testing. Upon intracerebroventricular treatment with scopolamine 30 min
- 10 prior to testing, the rats displayed significantly increased latencies in finding the platform, demonstrating deficits in learning. In rats treated with LVV-haemorphin-7 25 min after scopolamine, the scopolamine-induced latency in finding the platform was totally reversed, and these
- 15 rats were indistinguishable from the control group. Withdrawal of treatment on day 8 brought the latency of scopolamine-treated group back to control levels, indicating that the scopolamine-induced amnesia is reversible.

20

Example 13 **Effect of LVV-Haemorphin-7 on Acetylcholine Release in Rat Hippocampus**

Acetylcholine is thought to be the major transmitter involved in the processing of cognitive

25 function, since anti-cholinergic drugs induce memory deficit and confusion. In Alzheimer's disease, neuronal loss has been reported in cholinergic-rich areas, particularly in the septohippocampal pathway. Angiotensin AT₄ receptors were found in high abundance in the basal

30 nucleus of Meynert, in the CA2 and dentate gyrus of the hippocampus, and in somatic and autonomic preganglionic motoneurons of the monkey brain. This pattern of receptor distribution closely resembles that of cholinergic neurones, and suggests that the AT₄ receptors may be

35 associated with cholinergic pathways centrally. Moreover, as shown in Example 12 LVV-haemorphin-7 can reverse the learning deficit induced by scopolamine (a muscarinic

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receptor antagonist). We therefore postulate that LVV-haemorphin-7 can modulate acetylcholine release from the septohippocampal neurones via the AT_4 receptors.

Rats are anaesthetized with sodium

5 pentobarbitone, and stereotaxically implanted with intracerebral guide cannulae either in the dorsal hippocampus (coordinates 3.8mm caudal to bregma, 2.5mm lateral to midline, and 3.0mm ventral to the surface of the skull) or ventral hippocampus (coordinates 5.3mm caudal to
10 bregma, 5.4mm lateral to midline, and 6.5mm ventral to surface of the skull). The guide cannulae are secured with dental cement anchored to three screws in the skull. Dummy probes are then inserted into the guide cannulae to prevent blockade of the cannulae. The rats are allowed to recover
15 for 5-7 days. On the day of the experiment, a microdialysis probe, with a 3 mm dialysis membrane, is inserted through the guide cannula and perfused with artificial cerebrospinal fluid (148 mM NaCl, 3 mM KCl, 1.4 mM CaCl, 0.8 mM MgCl, 1.3 mM NaH_2PO_4 , 0.2 mM Na_2HPO_4 , pH 7.4) at a
20 flow rate of 2.0 μ l/min. Neostigmine (1.0 μ M) is added to the artificial cerebrospinal fluid to facilitate recovery of acetylcholine. Four 20-min baseline samples are collected 1 h after probe insertion, followed by four 20-min samples during the experimental period when LVV-haemorphin-7 (1 μ mol dissolved in artificial cerebrospinal
25 fluid and 1 μ M neostigmine) is perfused through the probe. During the recovery period, the peptide perfusion is withdrawn and four 20-min samples are collected.

Acetylcholine in the dialysates is measured by
30 HPLC with electrochemical detection. Acetylcholine and choline are separated on a 10 cm polymer-based analytical column, and then converted to hydrogen peroxide and betaine by an immobilized enzyme reactor (acetylcholinesterase and choline oxidase) coupled to the analytical column. The
35 mobile phase consists of 35 mM sodium phosphate at pH 8.5 supplemented with the antibacterial reagent Kathoon CG.

**Example 14 Detection of β -globin Sequences in Different
Neuronal Cell Lines by RT-PCR**

Total RNA is isolated from the following cell

5 lines:

(a) NG 108 rat glioma-neuroblastoma hybrid,

(b) SKNMC human neuroblastoma, and

(c) PC 12W rat pheochromocytoma. The total RNA is prepared as follows: 10^7 cells are homogenized in 4 ml of
10 4M guanidine thiocyanate, 25 mM sodium citrate and 0.05 % sodium dodecyl sulphate followed by sequential addition of 0.4 ml of 2 M sodium acetate pH 4.0, 4 ml water saturated phenol, and 0.8 ml of chloroform-isoamyl alcohol. The homogenate is mixed and cooled on ice for 15 min followed
15 by centrifugation at 2000 g for 15 min. The aqueous phase is removed and subjected to 2 phenol-chloroform extractions before RNA is precipitated by the addition of isopropanol.

The mRNAs are then subjected to RT-PCR. cDNA is synthesized from approximately 20 μ g of total RNA, using
20 reverse transcriptase and random hexamers. Ten percent of the cDNA product was amplified by PCR through 40 cycles, with each cycle consisting of denaturation at 94°C for 1 min, annealing of primers at 60°C for 1 min and primer extension at 72°C for 1 min, followed by a final 10 min
25 incubation at 72°C. The primers used were 5'CTGGTTGTCTACCCCTGGACTCAGAG3' (SEQ ID NO:2), and 5'CAGCACAACCACTAGCACATTGCC3' (SEQ ID NO:5), which corresponded with high homology to sheep β, δ, ϵ globin chains and flanked a 255 bp cDNA fragment. The sense primer
30 spans the nucleotide sequence which coded for LVV-haemorphin-7, and the antisense primer spans the second intron of the globin gene, to enable cDNA to be distinguished from contaminating genomic DNA. The PCR products are transferred to a Hybond N+ membrane by
35 downward Southern blotting in 0.4 M NaOH. The membrane is hybridized at 42°C in 5xSSC, 5xDenhardt's solution and 0.5% sodium dodecyl sulphate, with a 32 P end-labelled

- 34 -

oligonucleotide 5'CTCAGGATCCACATGCAGCTTATCACAG3' (SEQ ID NO:3), which is internal to the primers used for PCR and binds to β , δ and ϵ globin chains. After 12 h of hybridization, the filter is washed at 42°C in a buffer with a final stringency of 0.5xSSC and 0.1% sodium dodecyl sulphate.

We have mapped the distribution of AT₄ receptors in the brain of *Macaca fascicularis* and sheep spinal cord. The receptor has a striking and unique distribution, including motor- and sensory-associated regions and pathways and cholinergic cell bodies, including all motor nuclei in the brain stem and spinal cord. We have demonstrated that Ang IV inhibits neurite outgrowth in cultured embryonic chicken neurones, and that this peptide may therefore have a role in growth and development of the central and peripheral nervous systems.

We have purified an endogenous brain peptide which binds to the AT₄ receptor with high affinity. This decapeptide is 100% identical to the internal amino-acid sequence 30-39 of sheep β -globin. The presence of this β -globin-like sequence was demonstrated in sheep brain and other tissues using PCR. Screening of a rat brain cDNA library led to the isolation of a clone identical in sequence to rat β^A -globin.

We have demonstrated the presence of β -globin mRNA in brain tissue and isolated a β -globin cDNA clone from a rat brain library. These data suggest that LVV-haemorphin-7 is derived from β -globin precursors synthesized in the brain, although contamination by reticulocytes cannot be excluded. All of the cDNA clones sequenced correspond to the sequence encoding rat β^A -globin. The rat LVV-haemorphin-7 peptide sequence has a conservative substitution at position 10, with a tyrosine replacing a phenylalanine.

It therefore appears that a peptide corresponding to the sequence of the bovine LVV-haemorphin-7 exists in

- 35 -

brain, and is derived from β -globin as precursor. The peptide is almost certainly an endogenous ligand for abundant brain AT_4 receptors, and may therefore exert a range of actions on defined motor sensory and cholinergic
5 neurones.

We have shown that LVV-haemorphin-7 reverses the memory-disruptive effects of scopolamine in both passive avoidance conditioning trials and in water maze acquisition trials. However, administration of high doses of the
10 peptide may be detrimental to learning due to overstimulation of the neuronal system.

In a wider context, our findings suggest that β -globin may be a precursor of a range of neuroactive peptides generated in the central nervous system by
15 specific cleavage enzymes to interact with a range of receptors.

It will be apparent to the person skilled in the art that while the invention has been described in some
20 detail for the purposes of clarity and understanding, various modifications and alterations to the embodiments and methods described herein may be made without departing from the scope of the inventive concept disclosed in this invention.

25

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30 Harding, J.W.
***Neuroendocrinology*, 1995 16 23-52** ✓

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SEQUENCE LISTING

(1) GENERAL INFORMATION:

- 5
- (i) APPLICANT: HOWARD FLOREY INSTITUTE OF
EXPERIMENTAL PHYSIOLOGY AND MEDICINE
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CHAI, SIEW YEEN
10 MOELLER, INGRID
ALDRED, PETER
SMITH, IAN A
LEW, REBECCA A
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- (iii) NUMBER OF SEQUENCES: 7
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- 30 (A) MEDIUM TYPE: Floppy disk
(B) COMPUTER: IBM PC compatible
(C) OPERATING SYSTEM: PC-DOS/MS-DOS
(D) SOFTWARE: PatentIn Release #1.0, Version
#1.30
- (vi) CURRENT APPLICATION DATA:
- 35 (A) APPLICATION NUMBER: AU PO0893
(B) FILING DATE: 09-JUL-1996
(C) CLASSIFICATION:
- (viii) ATTORNEY/AGENT INFORMATION:
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- 39 -

(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:

- 5 (A) LENGTH: 10 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

10 (ii) MOLECULE TYPE: peptide

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

15 (v) FRAGMENT TYPE: internal

20 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

Leu Val Val Thr Pro Thr Thr Gly Ala Pro
1 5

10

25

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

- 30 (A) LENGTH: 26 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

35 (ii) MOLECULE TYPE: peptide

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

40 (v) FRAGMENT TYPE: internal

45 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

CTGGTTGTCT ACCCCTGGAC TCAGAG
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- 40 -

(2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 26 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: YES

(v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

CTCTGAGTCC AGGGGTAGAC AACCAG
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(2) INFORMATION FOR SEQ ID NO:4:

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(A) LENGTH: 28 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: YES

(v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

CTCAGGATCC ACATGCAGCT TATCACAG
28

- 41 -

(2) INFORMATION FOR SEQ ID NO:5:

- (i) SEQUENCE CHARACTERISTICS:
5 (A) LENGTH: 24 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
10
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: YES
- 15 (v) FRAGMENT TYPE: internal

20 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

CAGCACAACC ACTAGCACAT TGCC
24

25 (2) INFORMATION FOR SEQ ID NO:6:

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30 (A) LENGTH: 1244 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA
- 35 (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (v) FRAGMENT TYPE: not applicable
40

45 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

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TTGTGTTGAC 60

TCACAAACTC AGAAACAGAC ACCATGGTGC ACCTGACTGA TGCTGAGAAG
50 GCTGCTGTTA 120

- 42 -

ATGGCCTGTG GGGAAAGGTG AACCCCTGATG ATGTTGGCTG ATGCTGAGAA
 GGCTGCTGTT 180

5 AATGGCCTGT GGGGAAAGGT GAACCCTGAT GATGTTGGTG GCGAGGCCCT
 GGGCAGGCTG 240

CTGGTTGTCT ACCCTTGGAC CCAGAGGTAC TTTGATAGGT GGCGAGGCCC
 TGGGCAGGCT 300

10 GCTGGTTGTC TACCCTTGGA CCCAGAGGTA CTTTGATAGC TTTGGGGACC
 TGTCTCTGCTG 360

CTCTGCTATC ATGGGTAACC CTAAGGTGAA GGCCCATGGG CTTTGGGGAC
 CTGTCCTCTG 420

15 CCTCTGCTAT CATGGGTAAC CCTAAGGTGA AGGCCCATGG CAAGAAGGTG
 ATAAACGCCT 480

20 TCAATGATGG CCTGAAACAC TTGGACAACC TCAAGGGCAG GCAAGAAGGT
 GATAAACGCC 540

TTCAATGATG GCCTGAAACA CTTGGACAAC CTCAAGGGCA CCTTTGCTCA
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 GCAATATGAT 720

30 TGTGATTGTG TTGGGCCACC ACCTGGGCAA GGAATTCACC CGGGCTCCTG
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 35 CAGGCTGCCT 840

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40 TTCCAGAAGG TGGTGGCTGG AGTGGCCAGT GCCCTGGCTC ACAAGTACCA
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 50 TCTGTCAGTT 1140

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GTTGTCAAAA TGACAAAGAC CTTTGAAAAT CTGTCCTACT AATAAAAGGC
ATTTACTTTC 1200

5 ACTGCAAAAA AAAAAAAAAA AAAGAAGGCA TTTACTTTCA CTGC
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(A) LENGTH: 649 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

15 (ii) MOLECULE TYPE: DNA

(iii) HYPOTHETICAL: NO

20

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

25 CACAAACTCA GAAACAGACA CCATGGTGCA CCTGACTGAT GCTGAGAAGG
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30 GCTGCTGGTT GTCTACCCTT GGACCCAGAG GTACTTTGAT AGGAGTGCTT
TGGGGACCTG 180

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AGCAAGAAGG 240

35 TGATAAACGC CTTCAATGAT GGCCTGAAAC ACTTGGACAA CCTCAAGGGC
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40 GCTCATCTGA GTGAACTCCA CTGTGACAAG CTGCATGTGG ATCCTGAGAA
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CTCCTGGGCA ATATGATTGT GATTGTGTTG GGCCACCACC TGGGCAAGGA
ATTCACCCGG 420

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CACAAAAGAA AAGTACCACT AAACCTCTTT TCCTGCTCTT GTCTTTGTGC
AATGGTCAAT 540

50 TGTTCCTCAAG AGAGCATCTG TCAGTTGTTG TCAAAATGAC AAAGACCTTT
GAAAATCTGT 600

Sequence "GTTGTCAAAA"

- 44 -

CCTACTAATA AAAGGCATTT ACTTTCCTG CAAAAAAAAA AAAAAAAAAA
649

649 CCTACTAATA AAAGGCATTT ACTTTCCTG CAAAAAAAAA AAAAAAAAAA

CLAIMS

1. A method of modulating neuronal activity,
comprising the step of administering an effective amount of
5 a neuroactive peptide having at least one of the
biological activities of angiotensin IV as herein defined,
comprising the amino acid sequence:
Leu-Val-Val-Tyr-Pro-Trp-Thr-Gln-Arg-Phe, (SEQ ID NO:1),
or a biologically-active analogue or fragment of said
10 peptide, to a mammal in need of such treatment.
2. A method of modulating neuronal activity,
comprising the step of administering a biologically-active
non-peptide analogue of the neuronal peptide according to
claim 1 to a mammal in need of such treatment.
- 15 3. A method according to claim 2, in which the
biologically-active analogue is a peptidomimetic compound.
4. A method according to any one of claims 1 to 3,
in which the biological activity is selected from the group
consisting of modifying learning, modifying behaviour,
20 vasoactive effects, dilation of cerebral arteries, increase
in renal blood flow, increase in stereotypy behaviour,
facilitating memory retrieval, neurite modelling and
alleviation of the effects of spinal cord injury.
5. A method according to any one of claims 1 to 4,
25 wherein said neuronal activity is selected from the group
consisting of motor neuron activity, cholinergic neuron
activity and neuronal development.
6. A method of treating a condition selected from
the group consisting of dementia; Alzheimer's disease;
30 neuro-degenerative disorders involving one or more of
cholinergic pathways, motor pathways, or sensory pathways;
motor neuron disease; sensory peripheral neuropathies;
motor peripheral neuropathies; brain injury; and spinal
cord injury resulting from one or more trauma, hypoxia, and
35 vascular disease, comprising the step of administering an
effective amount of a neuroactive peptide having at least
one of the biological activities of angiotensin IV as

herein defined, comprising the amino acid sequence:
Leu-Val-Val-Tyr-Pro-Trp-Thr-Gln-Arg-Phe, (SEQ ID NO:1),
or a biologically-active analogue or fragment of said
peptide, to a mammal in need of such treatment.

- 5 7. A method according to claim 6, comprising the
step of administering a biologically-active non-peptide
analogue of the neuroactive peptide of claim 6 to a subject
in need of such treatment.
8. A method according to claim 7, in which the
10 biologically-active analogue is a peptidomimetic compound.
9. A method according to any one of claims 6 to 8,
in which the biological activity is selected from the group
consisting of modifying learning, modifying behaviour,
vasoactive effects, dilation of cerebral arteries, increase
15 in renal blood flow, increase in stereotypy behaviour,
facilitating memory retrieval, neurite modelling and
alleviation of the effects of spinal cord injury.
10. A method according to any one of claims 1 to 9,
in which the mammal is a human.
- 20 11. A method of screening for putative agonists or
antagonists of the effect of LVV-haemorphin-7 on neuronal
activity, comprising the step of testing the ability of the
compound to stimulate or inhibit the effect of LVV-
haemorphin-7 on a biological activity selected from the
25 group consisting of modifying learning, modifying
behaviour, vasoactive effects, dilation of cerebral
arteries, increase in renal blood flow, increase in
stereotypy behaviour, facilitating memory retrieval,
neurite modelling and alleviation of the effects of spinal
30 cord injury.
12. An antagonist of LVV-haemorphin-7, identified by
the method of claim 11.
13. An agonist of LVV-haemorphin-7, identified by the
method of claim 11.
- 35 14. A method of modulating neuronal activity,
comprising the step of administering an effective amount of
an antagonist according to claim 11 to a mammal in need of

such treatment.

15. A method of modulating neuronal activity,
comprising the step of administering effective amount of an
agonist according to claim 12 to a mammal in need of such
5 treatment.
16. A pharmaceutical composition comprising an
agonist according to claim 11, together with a
pharmaceutically acceptable carrier.
17. A pharmaceutical composition comprising an
10 antagonist according to claim 12, together with a
pharmaceutically acceptable carrier.

ABSTRACT

The invention relates to neuroactive peptides or analogues thereof, having at least one of the biological activities
5 of angiotensin IV, and which comprise the sequence Leu-Val-Val-Tyr-Pro-Trp-Thr-Gln-Arg-Phe, to methods of modulating neuronal activity, and to pharmaceutical composition thereof.

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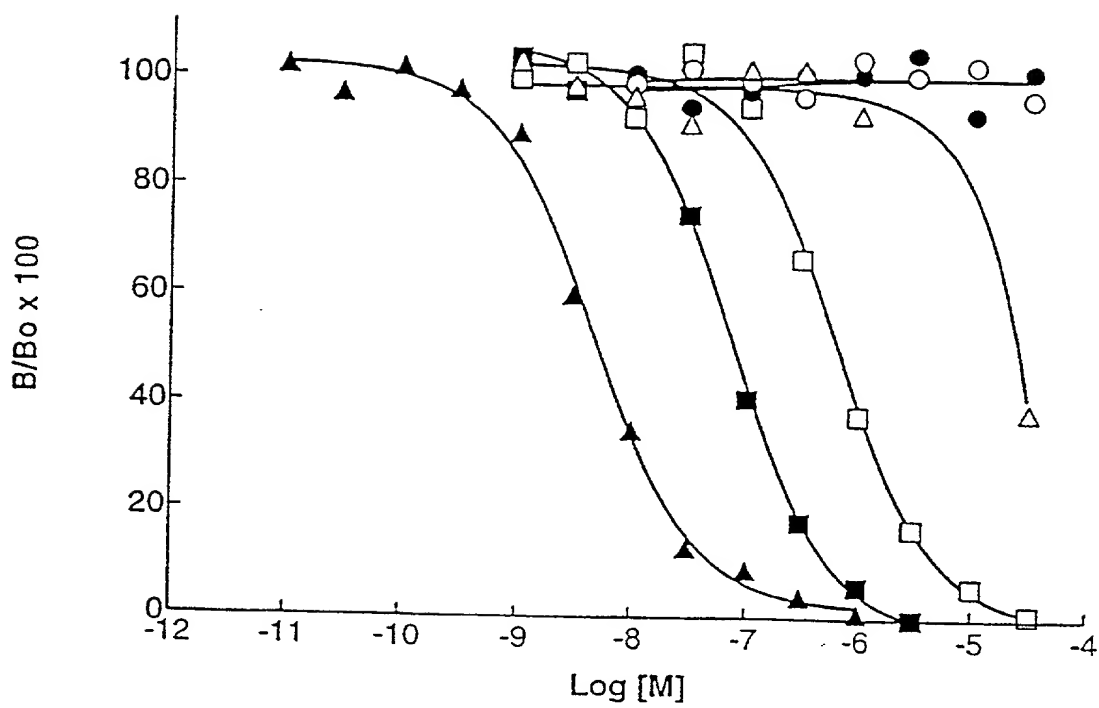


Figure 1

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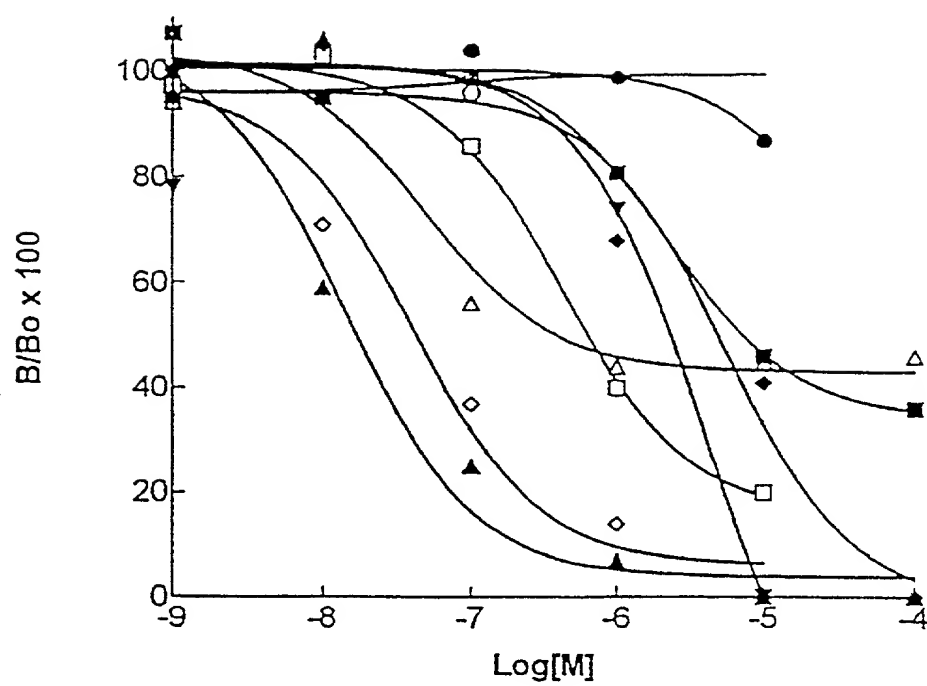


Figure 2

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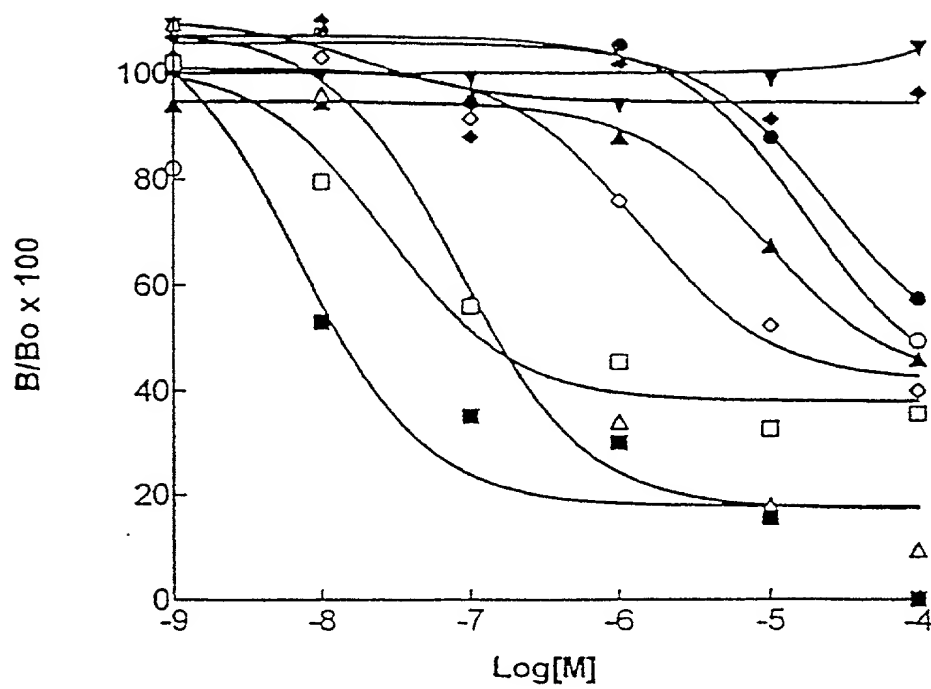


Figure 3

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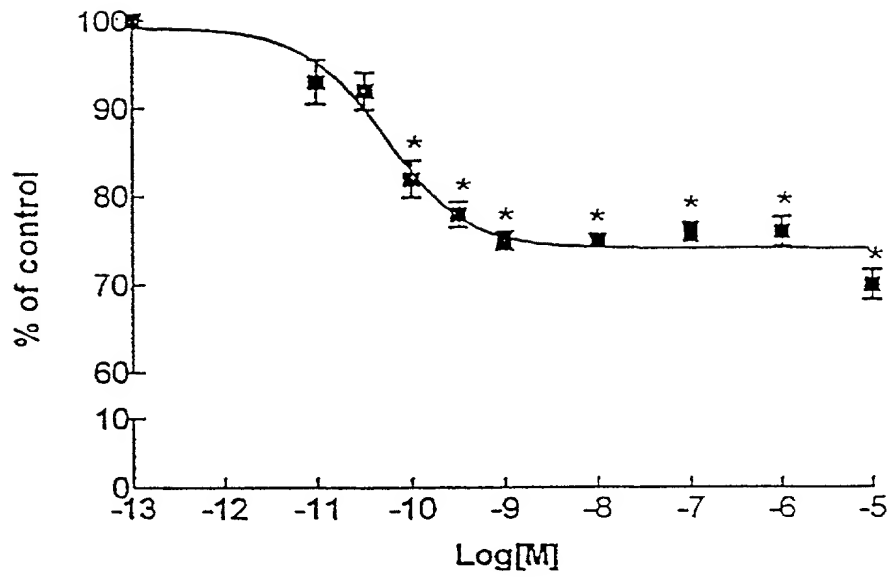


Figure 4

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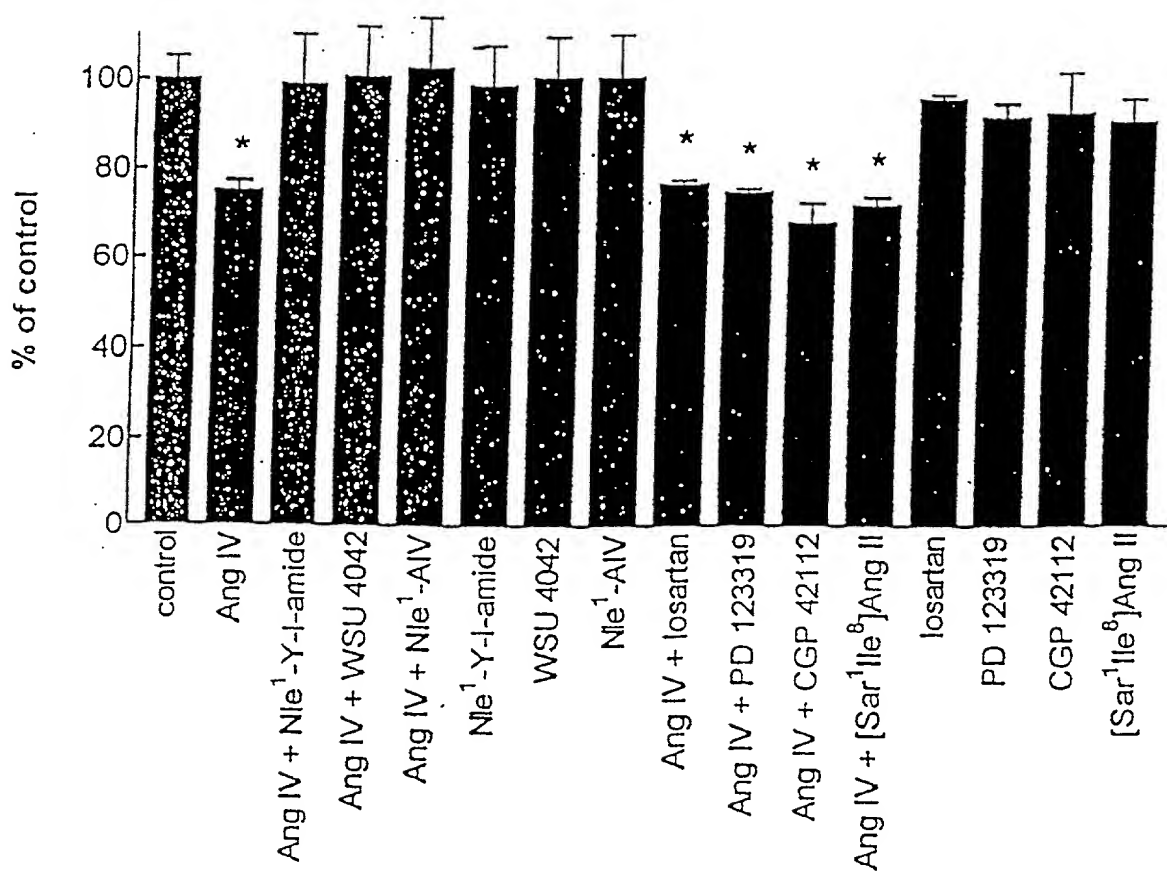


Figure 5

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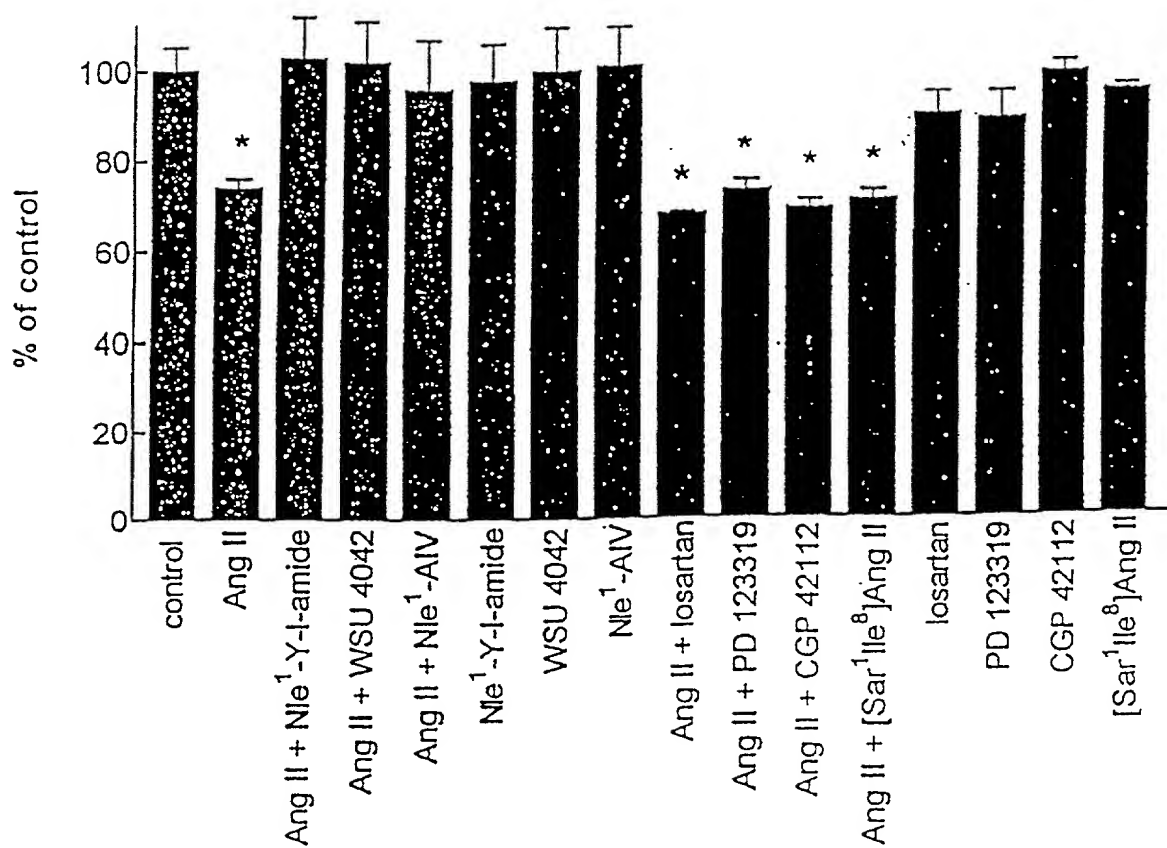


Figure 6

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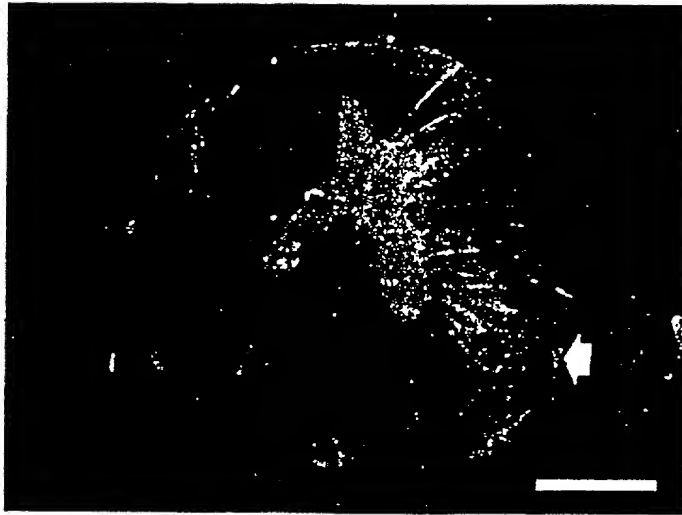


Figure 7

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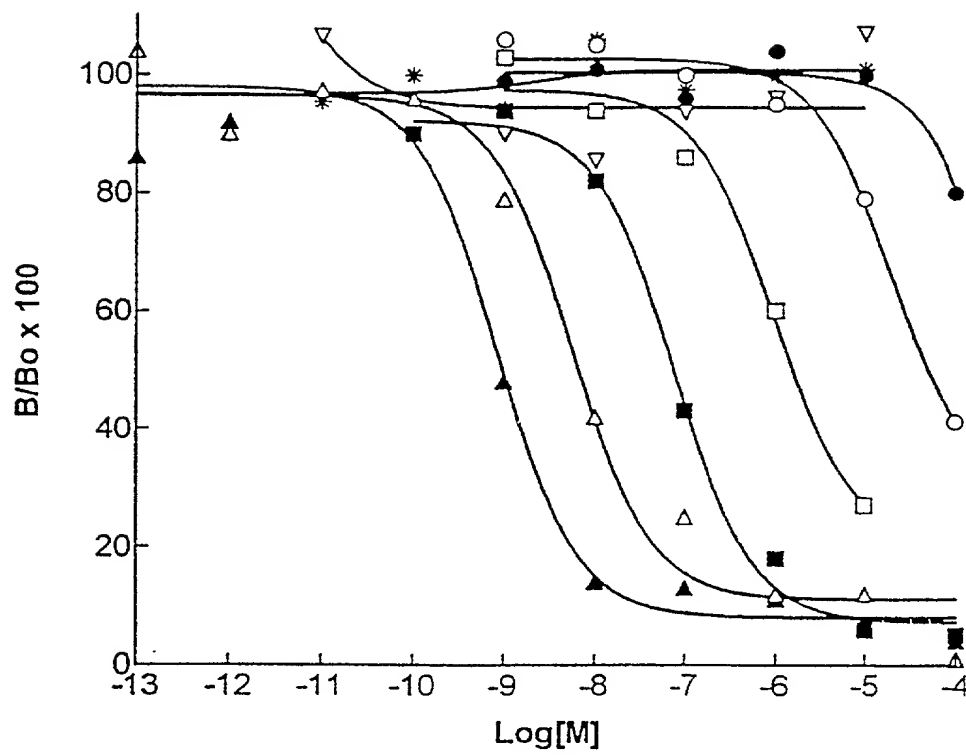


Figure 8

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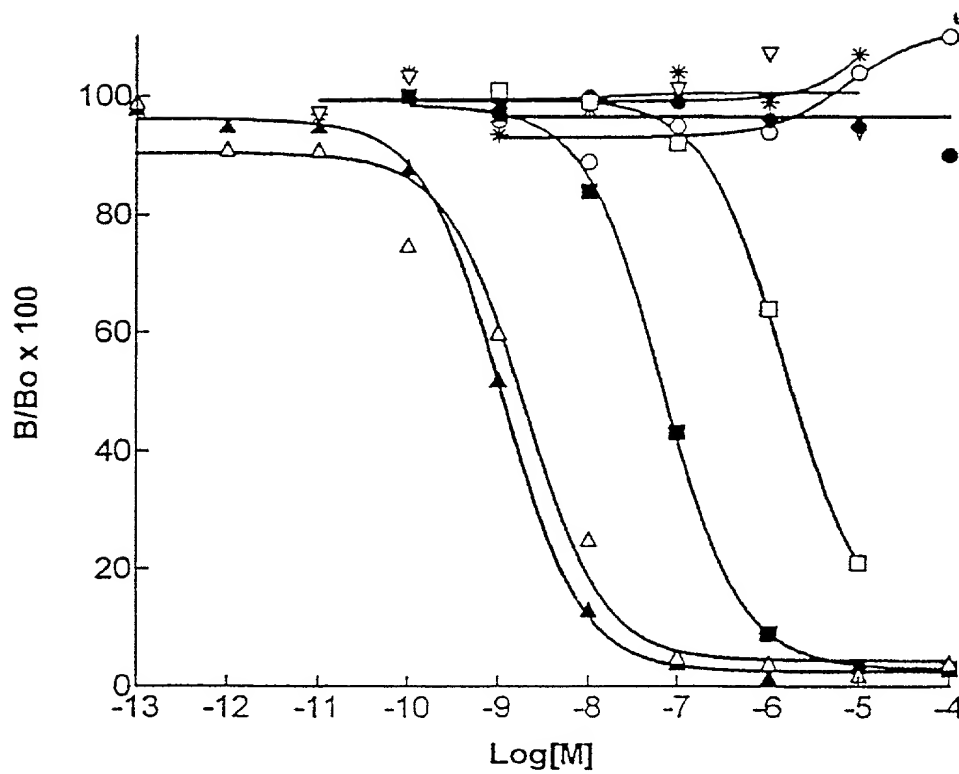
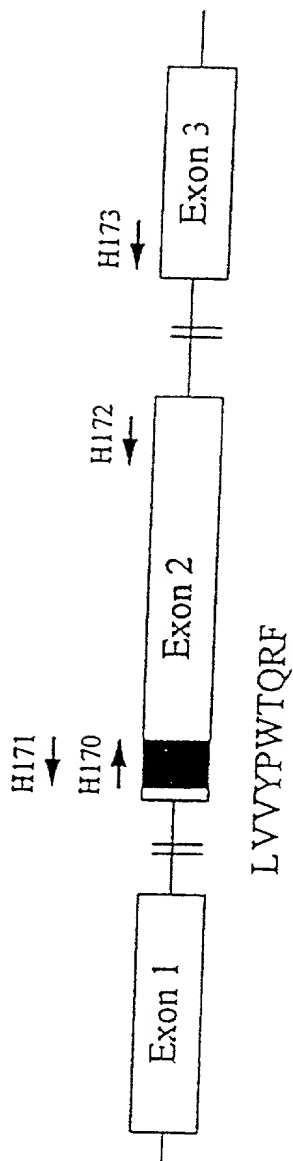


Figure 9

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 β - Globin Gene Structure

A



B

Oligonucleotide sequences:

H170: 5' CTGGTTGCTACCCCTGGACTCAGAG 3'
 H171: 5' CTCTGAGTCCAGGGGTAGACACCAG 3'
 H172: 5' CTCAGGATCCACATGCAGCTTATCAGAG 3'
 H173: 5' CAGCACACCCTAGCACATTGCC 3'

Figure 10

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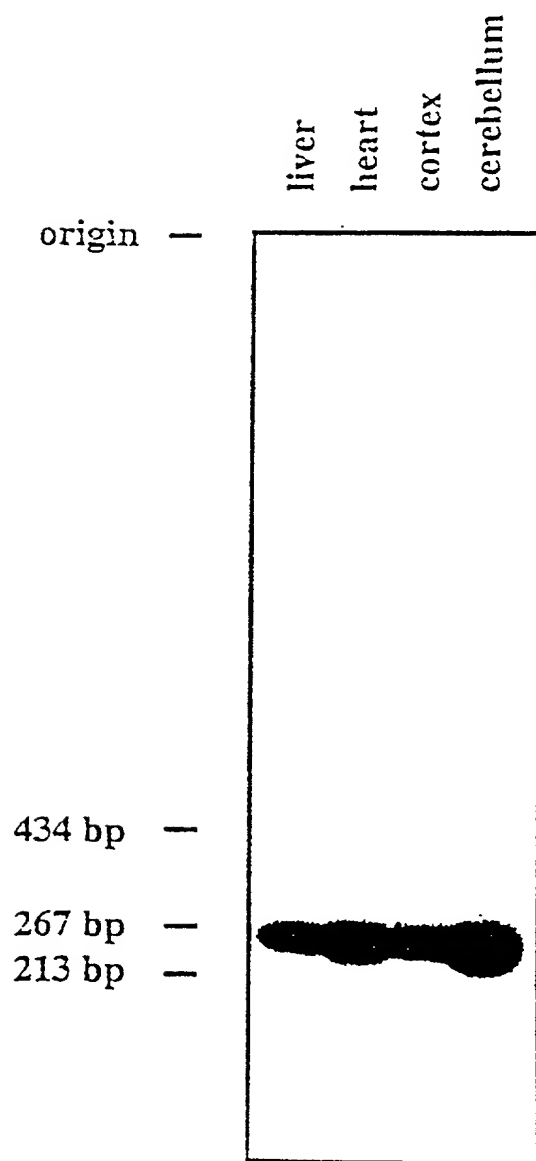


Figure 11

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EX				10	20	30
	CACAAACTCAGAAACAGACACCATGGTGCACCTGA					
	::					
RNBGLO	TGCTTCTGACATAGTTGTGTTGACTCACA	AACTCAGAAACAGACACCATGGTGCACCTGA				
	10	20	30	40	50	60
EX	40	50	60	70	80	90
	CTGATGCTGAGAAGGCTGCTGTTAATGGCCTGTGGG	GAAAGGTGAACCCTGATGATGTTG				
	::					
RNBGLO	CTGATGCTGAGAAGGCTGCTGTTAATGGCCTGTGGG	GAAAGGTGAACCCTGATGATGTTG				
	70	80	90	100	110	120
EX	100	110	120	130	140	150
	GTGGCGAGGCCCTGGGCAGGCTGCTGGTTGTCTACCCTTGGACCCAGAGGTACTTTGATA					
	::					
RNBGLO	GTGGCGAGGCCCTGGGCAGGCTGCTGGTTGTCTACCCTTGGACCCAGAGGTACTTTGATA					
	120	140	150	160	170	180
EX	160	170	180	190	200	210
	GCTTTGGGGACCTGTCCTCTGCCTCTGCTATCATGGGTAACCCTAAGGTGAAGGCCCATG					
	::					
RNBGLO	GCTTTGGGGACCTGTCCTCTGCCTCTGCTATCATGGGTAACCCTAAGGTGAAGGCCCATG					
	190	200	210	220	230	240
EX	220	230	240	250	260	270
	GCAAGAAGGTGATAAACGCCTTCAATGATGGCCTGAAACACTTGGACAACCTCAAGGGCA					
	::					
RNBGLO	GCAAGAAGGTGATAAACGCCTTCAATGATGGCCTGAAACACTTGGACAACCTCAAGGGCA					
	250	260	270	280	290	300
EX	280	290	300	310	320	330
	CCTTTGCTCATCTGAGTGAAGTCCACTGTGACAAGCTGCATGTGGATCCTGAGAACTTCA					
	::					
RNBGLO	CCTTTGCTCATCTGAGTGAAGTCCACTGTGACAAGCTGCATGTGGATCCTGAGAACTTCA					
	310	320	330	340	350	360
EX	340	350	360	370	380	390
	GGCTCCTGGGCAATATGATTGTGATTGTGTTGGGCCACCACTGGGCAAGGAATTACCC					
	::					
RNBGLO	GGCTCCTGGGCAATATGATTGTGATTGTGTTGGGCCACCACTGGGCAAGGAATTACCC					
	370	380	390	400	410	420
EX	400	410	420	430	440	450
	CCTGTGCACAGGCTGCCTTCCAGAAGGTGGTGGCTGGAGTGGCCAGTGCCTGGCTCACA					
	::					
RNBGLO	CCTGTGCACAGGCTGCCTTCCAGAAGGTGGTGGCTGGAGTGGCCAGTGCCTGGCTCACA					
	430	440	450	460	470	480
EX	460	470	480	490	500	510
	AGTACCACTAAACCTCTTTTCTGCTCTTGTCTTTGTGCAATGGTCAATTGTTCCCAAGA					
	::					
RNBGLO	AGTACCACTAAACCTCTTTTCTGCTCTTGTCTTTGTGCAATGGTCAATTGTTCCCAAGA					
	490	500	510	520	530	540
EX	520	530	540	550	560	570
	GAGCATCTGTCAGTTGTTGTCAAAATGACAAAGACCTTTGAAAATCTGTCCTACTAATAA					
	::					
RNBGLO	GAGCATCTGTCAGTTGTTGTCAAAATGACAAAGACCTTTGAAAATCTGTCCTACTAATAA					
EX	580	590	600	610		
	AAGGCATTTACTTTCACTGCAAAAAAAAAAAAAAAAAA					
	::::::::::::::::::::::::::::::::					
RNBGLO	AAGGCATTTACTTTCACTGC					

Figure 12

[illegible]

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EX CACAAACTCAGAAACAGACACCATGGTGCACCTGA
M V H L

EX CTGATGCTGAGAAAGGCTGCTGTTAATGGCCTGTGGGGAAAGGTGAACCCTGATGATGTTG
T D A E K A A V N G L W G K V N P D D V

EX GTGGCGAGGCCCTGGGCAGGCTGCTGGTTGTCTACCCTTGGACCCAGAGGTACTTTGATA
G G E A L G R L L V V Y P W T Q R Y F D

EX GCTTTGGGGACCTGTCCTCTGCCTCTGCTATCATGGGTAACCCTAAGGTGAAGGCCCATG
S F G D L S S A S A I M G N P K V K A H

EX GCAAGAAGGTGATAAACGCCCTTCAATGATGGCCTGAAACACTTGGACAACCTCAAGGGCA
G K K V I N A F N D G L K H L D N L K G

EX CCTTTGCTCATCTGAGTGAACCTCCACTGTGACAAGCTGCATGTGGATCCTGAGAACTTCA
T F A H L S E L H C D K L H V D P E N F

EX GGCTCCTGGGCAATATGATTGTGATTGTGTTGGGCCACCACTGGGCAAGGAATTCACCC
R L L G N M I V I V L G H H L G K E F T

EX CCTGTGCACAGGCTGCCTTCCAGAAGGTGGTGGCTGGAGTGGCCAGTGGCCTGGCTCACA
P C A Q A A F Q K V V A G V A S A L A H

EX AGTACCACTAAACCTCTTTTCCTGCTCTTGTCTTTGTGCAATGGTCAATTGTTCCCAAGA
K Y H *

EX GAGCATCTGTCAGTTGTTGTCAAAATGACAAAGACCTTTGAAAATCTGTCCTACTAATAA

EX AAGGCATTACTTTCACTGCAAAAAAAAAAAAAAAAAA

Figure 13

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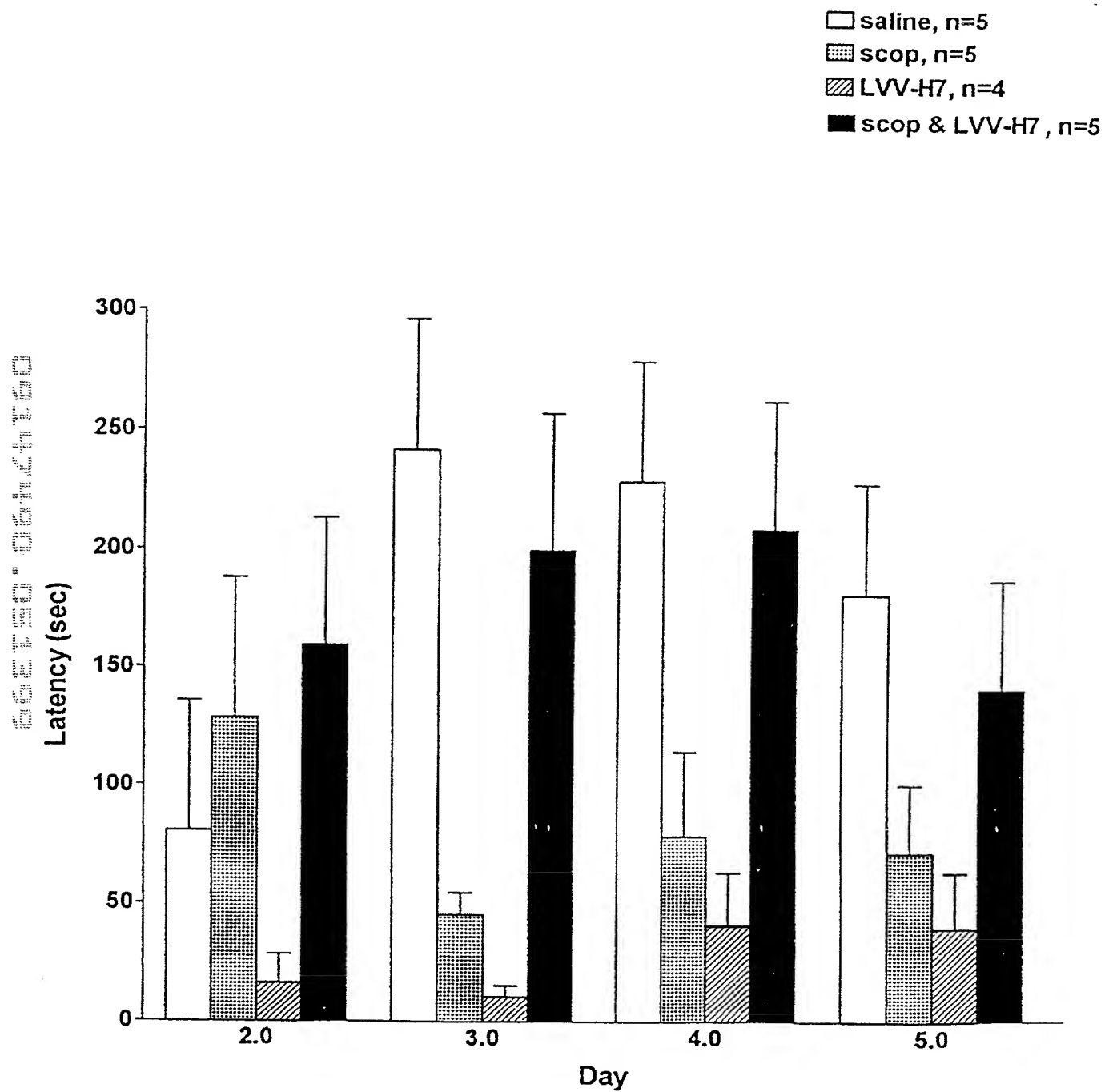


Figure 14

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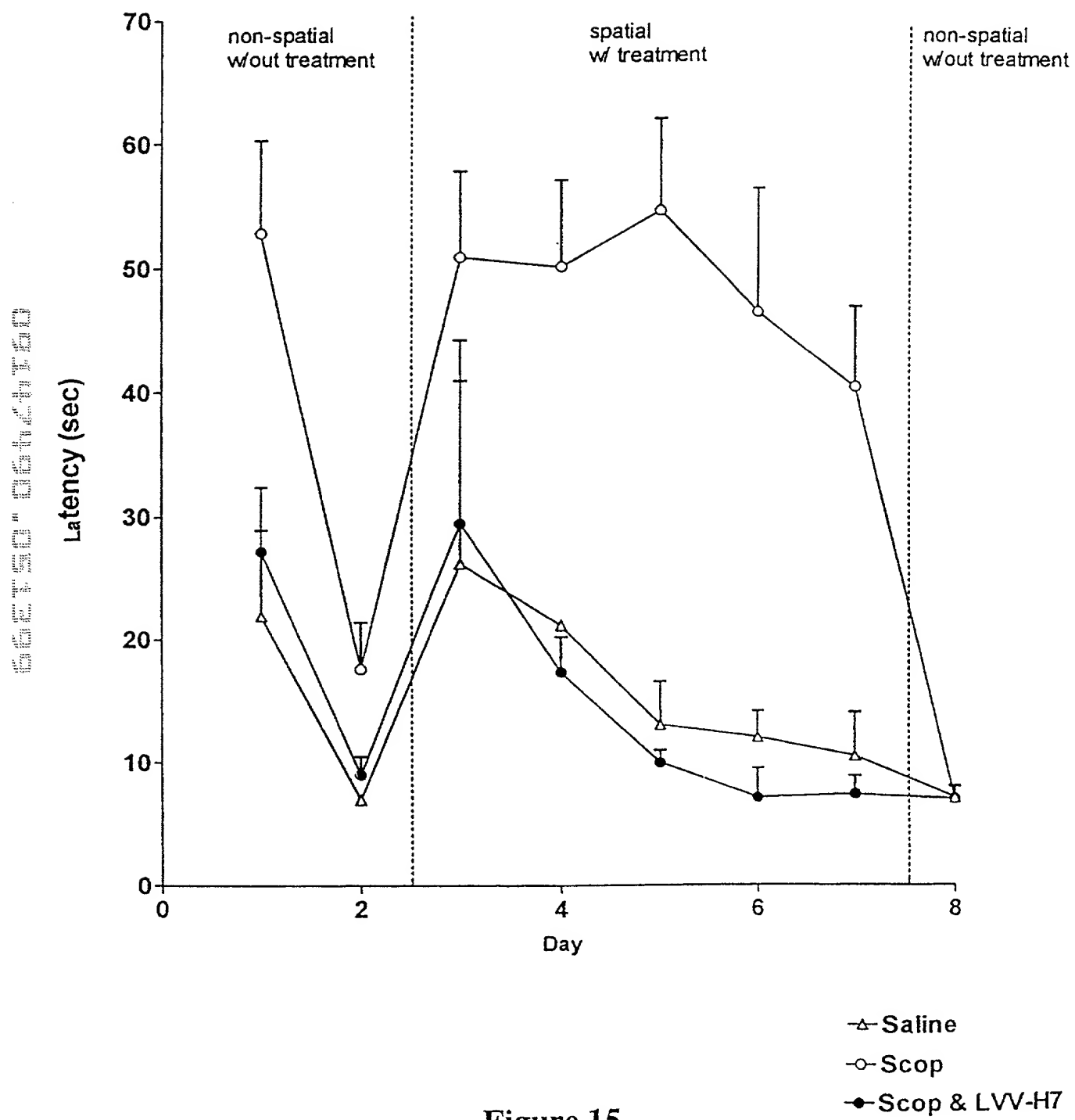


Figure 15

DECLARATION AND POWER OF ATTORNEY

As a below named inventor, I hereby declare that:

My residence, post office address, and citizenship are as stated below next to my name.

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled:

NEUROACTIVE PEPTIDE

the specification of which is annexed hereto unless the following box is checked:

☒ was filed on July 2, 1999 as United States Application Number or PCT International Application Number PCT/AU97/00436 and was amended on (if applicable).

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information which is known by me to be material to patentability as defined in Title 37, Code of Federal Regulations § 1.56.

I hereby claim foreign priority benefits under Title 35, United States Code, § 119(a)-(d) or § 365(b) of any foreign application(s) for patent or inventor's certificate, or § 365(a) of any PCT International application which designated at least one country other than the United States, listed below and have also identified below any foreign application for patent or inventor's certificate, or PCT International application having a filing date before that of the application on which priority is claimed:

PRIOR FOREIGN APPLICATION(S)

NUMBER	COUNTRY	DAY/MONTH/YEAR FILED	PRIORITY CLAIMED
PO 0893	Australia	9/July/1996	Yes

I hereby claim the benefit under Title 35, United States Code § 119(e) of any United States provisional application(s) listed below.

APPLICATION NO.	FILING DATE

I hereby claim the benefit under Title 35, United States Code, § 120 of any United States application(s), or § 365(c) of any PCT International application designating the United States, listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States or PCT International application in the manner provided by the first paragraph of Title 35, United States Code, § 112, I acknowledge the duty to disclose information which is known by me to be material to patentability as defined in Title 37, Code of Federal Regulations § 1.56 which became available between the filing date of the prior application and the national or PCT International filing date of this application:

APPLICATION SERIAL NO.	FILING DATE	STATUS: PATENTED, PENDING, ABANDONED

I hereby appoint as my attorneys, with full powers of substitution and revocation, to prosecute this application and transact all business in the Patent and Trademark Office connected therewith: Stephen A. Bent, Reg. No. 29,753; David A. Blumenthal, Reg. No. 26,257; William T. Ellis, Reg. No. 26,874; John J. Feldhaus, Reg. No. 28,822; Patricia D. Granados, Reg. No. 33,583; John P. Isacson, Reg. No. 33,715; Donald D. Jeffery, Reg. No. 19,980; Eugene M. Lee, Reg. No. 32,039; Richard Linn, Reg. No. 25,144; Peter G. Mack, Reg. No. 26,001; Brian J. McNamara, Reg. No. 32,789; Sybil Meloy, Reg. No. 22,749; George E. Quillin, Reg. No. 32,792; Colin G. Sandercock, Reg. No. 31,298; Bernhard D. Saxe, Reg. No. 28,665; Charles F. Schill, Reg. No. 27,590; Richard L. Schwaab, Reg. No. 25,479; Arthur Schwartz, Reg. No. 22,115; Harold C. Wegner, Reg. No. 25,258.

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I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

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